



Variations in Vegetation Structure, Species Dominance and Plant Communities in South of the Eastern Desert-Egypt

Fawzy SALAMA^{1*}, Monier ABD EL-GHANI², Mohamed GADALLAH¹, Salah EL-NAGGAR¹, Ahmed AMRO¹

> ¹ Assiut University, Faculty of Science, Botany Department, Assiut University Street, Assiut 71516, Egypt; mohamed.aag2@yahoo.com; salahelnaggar40@yahoo.com; ahmed.amro81@yahoo.com; fawzy_salama2010@yahoo.com (*corresponding author)
> ² Cairo University, Faculty of Science, Botany Department, 12613 Giza, Cairo, Egypt; elghani@yahoo.com

Abstract

For two successive years, the floristic diversity and vegetation composition in the southern part of the Eastern Desert of Egypt were investigated through four transects (3 crossing the Eastern Desert and one along the Red Sea). The data collected from 142 stands covering the study area included the species composition, functional groups, chorology and occurrences (Q-values). A total of 94 plant species belonging to 33 different families were recorded, with Asteracea, Zygophyllaceae, Fabaceae,

Poaceae, Chenopodiaceae and Brassicaceae as the largest families. Shrubs represented the largest functional group (39.4%), while perennial herbs represented the smallest ones (12.8%). Species occurrence (Q-value) revealed that Zilla spinosa, Acacia tortilis subsp. raddiana, Morettia philaeana, Caroxylon imbricatum, Zygophyllum coccineum and Citrullus colocynthis had wide ecological range of distribution (dominant species, Q-values ≥ 0.2). Saharo-Arabian chorotype was highly represented (72.6%) in the flora of this area, eventually as mono, bi or pluriregional. Classification of the data set yielded 7 vegetation groups included: (A) Zilla spinosa-Morettia philaeana, (B1) Zilla spinosa-Citrullus colocynthis-Morettia philaeana, (B2) Zilla spinosa, (C1) Zygophyllum album-Tamarix nilotica, (C2) Zygophyllum coccineum-Tamarix nilotica, (D1) Zilla spinosa-Zygophyllum coccineum and (D2) Zilla spinosa-Acacia tortilis subsp. raddiana-Tamarix aphylla-Balanites aegyptiaca. Certain vegetation groups were assigned to one or more transects. Detrended Correspondence Analysis (DCA) revealed that electrical conductivity, sodium, potassium, calcium, magnesium, chlorides, moisture content, sulphates, pH, organic matter and gravel were the soil variables that affect the species distribution in this study.

Keywords: arid environment, chorology, flora, plant functional groups, soil-vegetation relationships.

Introduction

The Eastern Desert of Egypt occupies about 223,000 km², i.e. 21% of the total area of Egypt. It is characterized by two main ecological units, the Red Sea costal land and the inland desert with its wadis. In Egypt, like in the other arid lands, the desert vegetation is characterized by openness and is composed of a permanent framework of perennials, the interspaces of which may be occupied by ephemerals and their duration depends on the irregular rainfall and soil thickness (Zahran and Willis, 2009). The vegetation composition and plants distribution covering the Egyptian Eastern Desert and Red Sea coast such as in the northern inland part (Abd El-Ghani, 1998; Fossati et al., 1998), in Wadi Állaqi (Sheded *et al.*, 2012), in Wadi Degla (Hegazy et al., 2012), in Wadi Gimal (Galal and Fahmy, 2011; Gomaa, 2012), in Wadi Natash (Suzan et al., 2013) and in central Eastern Desert (Abd El-Ghani et al., 2013a; Salama and El-Naggar, 1991; Salama and Fayed, 1990;

Salama et al., 2012, 2013) have been studied.

Ecologically, this desert is characterized by two main units, the Red Sea costal land and the inland desert. The Red Sea coastal land extends from Suez to Mersa Halaib at the Sudano-Egyptian border, while the inland part lies between the range of the Red Sea coastal land in the east and the Nile Valley in the west (Hassib, 1951). It is a rocky plateau dissected by a number of wadis; each has a main channel with numerous tributaries.

The floristic compositions, life forms and chorological affinities of deserts paid the attention to many others. Along an altitudinal gradient in East Ladakh (NW Himalayas) of India, Klimes (2003) indicated the preponderance of hemicryptophytes followed by therophytes. Also, the main life-form classes in Brazilian (Cerrado sites) were the hemicryptophytes and the phanerophytes (Batalha and Martins, 2002). Therophytes also dominated many arid and semi-arid study areas as in northeastern Brazil (Da Costa *et al.*, 2007), in Mount Hymettus of central Greece (Gouvas and Theodoropoulos, 2007), in Khulais region of western Saudi Arabia (Al-Sherif *et al.*, 2013) and in Alborz Mountains of Iran (Mahdavi *et al.*, 2013).

One approach to addressing the complexity of desert vegetation is functional analysis. Plant species can be classified into functional groups based on a variety of characteristics. Functional groups have been defined as sets of species showing either similar responses to the environment or similar effect on major ecosystem processes (Gitay and Noble, 1997). Each functional group potentially will partition the environmental gradient differently (Austin, 1990; Dale, 1998; Lyon and Sagers, 2003; Smith and Huston, 1989).

The availability of water is one of the major factors reglating the species distribution in arid zones that are usually characterized by irregular, scanty and unpredictable rainfall, and long periods of drought (Noy-Meir, 1973; Yair and Danin, 1980). Soil-vegetation relationships were the subject of several investigations in the arid regions of the Middle East, e.g., Hillel and Tadmor (1962), Kassas and Girgis (1965), Olsvig-Whittaker et al. (1983), Stahr et al. (1985). Recently, multivariate analysis techniques were extensively used to elucidate these relationships, e.g., Moustafa and Zaghloul (1996), Abd El-Ghani (2000) Abd El-Ghani and Amer (2003), Abd El-Ghani et al. (2013 a, b), Salama et al. (2012, 2013).

This study aimed to (1) determine the spatial distribution patterns of the recorded species in terms of plant functional groups, (2) assess the soil factors which controlle the vegetation and to identify the regional plant communities, and (3) analyze the floristic variations between the northern and southern parts of the Eastern Desert.

Materials and methods

Study area

The study area covered nearly the southern quarter of the Eastern Desert (about 54,500 km²) between 26°45' and 24°1' N latitudes and 32°45' and 35°00' E longitudes (Fig. 1) It covered the area between Qena Governorate until Aswan Governorate on the Nile Valley and from Safaga until Berenice on the Red Sea coast. According to Zahran and Willis (2009), this area covered three desert types: (1) The limestone desert (Assiut-Qena Desert), (2) The sandstone desert (Idfu-Kom Ombo Desert), and (3) The Red Sea coastal plain. Detailed studies on the geology, geomorphology, topography and lithology have been documented by Said (1962), Abu Al-Izz (1971), and Zahran and Willis (2009).

The Egyptian desert is among the most arid parts of the world characterized by extreme aridity and high temperature. Available climatic records over the period 2003-2012 in four meteorological stations (Qena, Safaga, Aswan and Marsa Alam) demonstrated that the average monthly temperature ranged between 14.9°C in January (minimum) and 33.6°C (maximum) in July. Rainfall occurs only in winter and is due to random cloudbursts, a general feature in arid desert: rain may occur once every several years. Annual average rainfall records (over 30 years) showed notable decrease along north-south direction: 5.3 mm/year in Qene in the north along the Nile Valley to 3.4 mm/year in Quseir in the south along the Red Sea coast (Abd El-Ghani, 1998). Averages of relative humidity reached to the maximum of 51.5% and 52.7% (in December), while its minimum reached 25.6% and 32.4% (in June) for Mersa Alam and Safaga (Abd El-Ghani, 1998, Abd El-Ghani, 2013a).

Data collection and vegetation analysis

Between 2011 and 2012, vegetation sampling was performed in the study area using 4 transects representing the 3 desert types (Fig. 1). One hundred and forty-two georeferenced (using GPS model Garmin eTrex HC) randomly selected stands $(20 \times 30 \text{ m})$ were selected along the four transects to represent apparent variations in the physiognomy of vegetation and in the physiographic features. The sandstone desert included (T1) which comprised of Aswan-Berenice road (300 km; 24°05' - 24°00' N and 32°55' - 35°24' E); Wadi Kharit (250 km, 24°26' -24°12' N and 33°11' - 34°40' E); W. Natash (100 km, 24°21' - 24°40' N and 33°24' - 34°30' E), and W. Gimal (65 km, 24°34' - 24°40' N and 34°35' - 35°05' E); (T2) Idfu-Marsa Alam road (100 km, 25°55' N, 32°55' - 34°55' E). In the limestone desert, (T3) included Qena-safaga road (155 km, 26°12' - 26°46' N and 32°44' - 33°56' E), and along the Red Sea coastal plain (T4) that extends for about 240 km between 24°39' - 26°36' N and 32°05' - 34°00' E.

In this study, plants were categorized into four functional



Fig. 1. Location map of the study area, showing the distribution of the studied stands along the 4 transects

groups: trees, shrubs, perennial herbs and annual herbs. Correspondingly, in each transect plant species were segregated into four a priori defined functional groups: tree layer species, shrub layer species, and herb species. The presence/absence for each species was recorded in the studied stands, and a count-floristic list was obtained. The number of species within each functional group category was expressed as a percentage of total number of species in each transect. Analysis of phytogeographical ranges was carried out according to White and Léonard (1991). Taxonomic nomenclature and functional groups categorizing was according to Täckholm (1974), Boulos (1995, 1999, 2000, 2002, 2005) and El Hadidi and Fayed (1978, 1995). Voucher specimens of each species were collected, and identified at the Herbaria of Assiut University (ASTU) and Cairo University (CAI), where they were deposited.

The degree of occurrence of each species was determined using the Q-value (Danin *et al.*, 1985) as follows: Q= number of entries of a species X total number of species/13,348 (total number of entries). The Q-values and occurrences were categorized as follows: D=dominant, Q-value ≥ 0.2 ; VC=very common, Q-value 0.1– 0.199; C= common, Q-value 0.05-0.099; O=occasional or rare species, Q-value 0.01–0.049 and S= sporadic or very rare, Q-value ≤ 0.01 .

À floristic presence-absence data matrix of 142 stands and 94 species was subjected to classification by cluster analysis of the program Community Analysis Package (CAP) version 1.2 (Henderson and Seaby, 1999) using squared Euclidean distance dissimilarity matrix with minimum variance (also called Ward's method) as agglomeration criterion (Orlóci, 1978). The resulted vegetation groups (plant communities) were named after the dominant species that have the highest presences percentages in the stands of this group.

Detrended Correspondence Analysis (DCA) ordination based on species presence-absence data for each species was performed to examine patterns in species composition among species of different vegetation groups. The relationship between the vegetation and soil variables were assessed by calculating the simple linear correlation coefficient (r) between the DCA axes (reflect the vegetation gradient) and the soil variables.

One Way Analysis of Variance was applied to examine the statistical differences between the functional groups.

Soil sampling and analysis

Soil samples (0-50 cm depth) were collected at 3 random points from each stand as a profile (composite samples). These samples were then air-dried, thoroughly mixed, and pass through a 2 mm sieve to get rid of gravel and boulders. The weight of gravel in each stand was determined and expressed as a percentage of the total weight of the soil sample. The soil texture was determined using the sieve method; the amount of each fraction (sand, silt and clay) was expressed as percentage of the original weight used (Ryan *et al.*, 1996). Soil moisture content was estimated by drying at 105°C; then the percentage of soil moisture was calculated based on dry weight of the soil (Kapur and Govil, 2000). The soil

portion less than 2 mm in size was kept for chemical analysis according to Jackson (1967) and Allen and Stainer (1974). Soil water extracts (1:5) were prepared for determination of electrical conductivity (EC) using conductivity meter, and pH using a glass electrode pH-meter. Organic matter (OM) was determined using the Walkely and Black rapid titration (Black, 1979). Sodium and potassium were determined by flamephotometer. Calcium and magnesium were estimated by titration against EDTA (ethylenediamine dihydrogen tetraacetic acid) using ammonium purpurate and eriochrome black T as indicators (Jackson, 1967). Chlorides were determined by direct titration against AgNO₃ using potassium chromate as an indicator, and bicarbonates by direct titration against HCl using methyl orange as indicator. Sulphates were determined by a turbidemetric technique with barium chloride and acidic sodium chloride solution using spectrophotometer (Model 1200) according to Bardsley and Lancaster (1965).

Results

Floristic composition

In total, 94 species (62 perennials and 32 annuals) constituted the floristic composition, representing 76 genera and 33 families (Tab. 1). About more than 50% of these species belonged to 6 families arranged in the following sequence, *Asteraceae* > *Zygophyllaceae* > *Fabaceae* > *Poaceae* > *Chenopodiaceae* > *Brassicaceae*. The largest family was *Asteraceae* (7 genera and 10 species), while 18 families were monospecific. The total number of recorded species was 46, 35, 52 and 46 for T1, T2, T3 and T4, respectively.

In terms of functional groups (Fig. 2), it can be noted that trees and perennial herbs were the least (2-7 species) represented among the 4 studied transect, while annual herbs and shrubs were the most (14-24 species). The distribution of functional groups within the studied transects showed significant difference (F-value=3.11, P=0.032) for the Red Sea transect (T4) among the others (F-value= 0.92, P=0.44, F-value=0.51, P=0.68, and F-value=0.65, P=0.58 for T1, T2 and T3, respectively). Few grasses (*Poaceae*) were recorded within transects (5, 2, 1, 5 species in T1, T2, T3 and T4, respectively), whereas shrubs dominated (17, 14, 20 and 23 species in T1, T2, T3 and T4, respectively).

Tab. 1 showed the distribution of the different functional groups within the study area. The recorded 13 tress were, amongst others, *Acacia tortilis* subsp. *raddiana*, *Tamarix aphylla*, *Balanites aegyptiaca*, *Ziziphus spinachristi*, *Avicennia marina*, *Hyphaene thebaica*, and *Moringa peregrina*.

Shrubs were the largest (37 species) represented functional group. The widely distributed species included Zygophyllum coccineum, Zilla spinosa, Caroxylon imbricatum, Aerva javanica and Leptadenia pyrotechnica that occurred in all transects. Whereas Caroxylon villosum, Artemisia judaica, Atriplex leucoclada, Chrozophora oblongifolia, Fagonia mollis and F. bruguieri were represented only in the northern transect (T1), Zygophyllum album, Nitraria retusa, Limonium axillare, Arthrocnemum macrostachyum, Cornulaca monacantha, Taverniera aegyptiaca and Capparis spinosa were confined to the Red Sea transect (T4), and another 3 shrubs showed consistency to the southern sector (Aswan-Kharit-Gimal transect; T3).

Four perennial herbs (*Aeluropus littoralis, Juncus rigidus, Leptochloa fusca* and *Cyperus rotundus*) showed consistency to the Red Sea transect (T4). For the northern transect (T1), *Imperata cylindrica, Stipagrostis plumosa, Dichanthum annulatum* and *Typha domingensis* exhibited certain degree of consistency to this transect (Tab. 1).

Three annual herbs (Astragalus vogelii, Polycarpaea repens and Tetraena simplex) had wide range of distribution (occurred in all transects). The Aswan-Kharit-Gimal transect (T3) was characterized by Astragalus eremophilus, Hippocrepis constricta, Lupinus digitatus, Launaea amal-aminae, L. capitata, Polycarpaea robbairea and Glinus lotoides which were not recorded elsewhere (Tab. 1).

Species abundance

The recorded species were categorized according to their Q-values (Tab. 1) as follwos: (i) Dominant species, of which Zilla spinosa had presence value of 61%, and Acacia tortilis subsp. raddiana with P=36%. Caroxylon imbricatum and Zygophyllum coccineum (shrubs), Morettia philaeana (annual herb) and Citrullus colocynthis (perennial herb) had lower presence values; (ii) Very common species, 10 species (e.g., Tamarix aphylla, Fagonia thebaica, Aerva javanica, Pulicaria undulata, Schouwia purpurea); (iii) Common species, 20 species included some salt-tolerant species such as Nitraria retusa and Zygophyllum album, and Phragmites australis; (iv) Occasional species, constituted the main bulk of the flora (33 species, 35.1% of total species), with their Q-values ranged between 0.01 and 0.049; (v) Sporadic species, comprised of 25 species with Qvalues=0.007 which included 4 trees, 7 shrubs, 6 perennial herbs and 8 annual herbs.

Chorological affinities

The chorological spectrum of the recorded species was illustrated in Fig. 2. The cosmopolitan, palaeotropical and pantropical species constituted 12 species (12.8% of the total flora - Tab. 1). Monoregional Saharo-Arabian chorotype was well-represented (35 species) in the study area, while species of Sudano-Zambezian (*Crotalaria aegyptiaca*), Sudanian (*Acacia nilotica*), Mediterranean (*Lotus hebranicus* and *Lupinus digitatus*) and Irano-Turanian (*Cotula cinerea*) were very modestly represented.

A total of 30 species were bi-regional chorotypes representing 31.9% of the recorded species, distributed as follows: (1) 18 species belonging to Saharo-Arabian+Sudano-Zambezian (e.g., *Trichodesma africanum*, *Balanites aegyptiaca, Leptadenia pyrotechnica, Calotropis procera, Cleome amblyocarpa, Salvadora persica, Limonium* axillare and Hyphaene thebaica). (2) 7 species belonging to the Saharo-Arabian+Irano-Turanian (e.g., Tamarix aphylla, Launaea nudicaulis, Cleome droserifolia and Fagonia bruguieri, (3) 2 species belonging to Sudano-Zambezian+Guineo-Congo (Moringa peregrina and Oxystelma esculentum), (4) 1 species belonging to Mediterranean+Irano-Turanian (Aeluropus littoralis). (5) 2 species belonginig to Mediterranean+Saharo-(Panicum turgidum Arabian and Chrozophora oblongifolia). In general, 18 species were belonged to Saharo-Arabian+Sudano-Zambezian, while the Saharo-Arabian+Irano-Turanian species were represented by 7 species (Tab. 1).

About 12.8% of the recorded species (12 species) were pluri-regional with wide geographical range of distribution (e.g., *Citrullus colocynthis*, *Zygophyllum album*, *Arthrocnemum macrostachyum*, *Juncus rigidus* and *Capparis spinosa*).



Fig. 2. Chorotype spectrum and functional groups diagram of the study area. M= species magnitude and average group abundance

Classification of the vegetation

Application of classification using cluster analysis to the floristic presence-absence data matrix of the study area yielded 7 vegetation groups (Tab. 2, Fig. 3). Each of the identified vegetation group will be named after the dominant species (i.e., highest presence percentages). Notably, none of the recorded species occurred in all the identified groups. Apart from coarse sand, clay and bicarbonates, the other thirteen (out of total of 16) measured soil variables showed significant differences (p < 0.05, 0.01) between the vegetation groups (Tab. 3).

Group (A): Zilla spinosa-Morettia philaeana group

The 18 stands of this group (41 species) were mostly located along Idfu-Marsa Alam transect (T2), with soil soil rich in its organic matter (OM)content and and highest pH, but had the lowest contents of fine sand, water content, Mg⁺² and Cl. Co-dominant species included *Caroxylon imbricatum*, *Fagonia thebaica*, *Schouwia purpurea* and *Tetraena simplex*. Consistent species to this group were *Echium horridum*, *Glinus lotoides*, *Oxystelma esculentum*, *Caroxylon villosum*, *Stipagrostis plumosa* and *Tribulus megistopterus*.

Group (B1): Zilla spinosa-Citrullus colocynthis-Morettia philaeana group

The 18 stands of this group (26 species) were located along Wadi Natash, W. Kharit and El-Shekh El-Shazly-Marsa Alam road (T3). Soil contents of gravels, fine sand, OM and pH were higher than the total means. The lowest contents were recorded in Na⁺² and HCO₃. Beside the dominants, *Acacia tortilis* subsp. and *Senna italica* were the co-dominants. Some species were confined to this group such as *Chenopodium album* and *Filago desertorum*.

Group (B2): Zilla spinosa group

This group (7 stands, 30 species) was characterized by the dominance of Zilla spinosa (P=100%), distributed along (Aswan-Kharit-Gimal transect, T3). Most of the examined soil variables (gravels, clay, EC, OM, Na⁺, K⁺, Ca⁺², HCO₃⁻ and SO₄⁻²) attained their lowest levels in the stands of this group. However, fine sand content was the highest amongst the others. Among the important co-dominant species, Astragalus vogelii, Cotula cinerea and Launaea nudicaulis were included. Consistent species to this group were Launaea capitata and L. cassiniana.

Group (C1): Zygophyllum album-Tamarix nilotica group

Most stands of this group (41 stands, 32 species) were located along the Red Sea coast transect (T4) between Marsa Alam and Qusier, and occurred on saline soil with soluble anions and cations contents higher than the groups (A, B1, B2, D1 and D2). The dominant species of this group, together with the co-dominants *Nitraria retusa* and *Limonium axillare* exhibited the saline nature of this group. Certain species showed consistency to this group such as *Aeluropus littoralis, Arthrocnemum macrostachyum, Avicennia marina*.

Group (C2): *Zygophyllum coccineum-Tamarix nilotica* group

This group (8 stands) was the least diversified (19 species) among others. The stands of this group were mainly located in T4 (Qusier-Safaga transect) along the Red Sea coast occurred on saline soil with the highest silt, clay, electric conductivity, water content and all the examined ions. However, it recorded the lowest pH and coarse sand content. The co-dominant species included Phragmites australis, Nitraria retusa, Limonium axillare and Zygophyllum album. Four weed species (Chenopodium murale, Cyperus rotundus, Leptochloa fusca and Sonchus oleraceus) were recorded among the 6 confined species to this group.

Group (D1): Zilla spinosa-Zygophyllum coccineum group

This group of stands (31) was the most diversified (53 species) among other groups, and collected from three different transects (T1, T2 and T4) found on soil in rich

in gravels and poor in silt content. The other soil factors had intermediate position amongst the other groups. The co-dominant species included *Caroxylon imbricatum*, *Lotus hebranicus* and *Ochradenus baccatus*. Twelve species showed consistency to this group such as *Acacia nilotica*, *Moringa peregrina*, *Ziziphus spina-christi* (trees), *Atriplex leucoclada*, *Fagonia bruguieri* (shrubs), and *Dichanthum annulatum*, *Imperata cylindrica* (herbs).

Group (D2): Zilla spinosa-Acacia tortilis subsp.-Tamarix aphylla-Balanites aegyptiaca group

This group (19 stands, 20 species) was characterized by the combination of the dominant species, mostly located in Wadi Gimal and its tributaries (T3) on a soil rich in fine sand, silt, pH and K⁺ and poor in Mg⁺² and water contents. The co-dominants of this group had low presence values such as *Zygophyllum coccineum*, *Pulicaria undulata* and *Calotropis procera*. Two species were confined to this group, *Capparis decidua* and *Salvadora persica*.



Fig. 3. Dendrogram showing cluster analysis of the studied 142 stands, with the 7 vegetation groups (A-D2) separated.

Ordination of the vegetation

Analysis of 142 stands along axes 1 and 2 (eigenvalues 0.707 and 0. 497, respectively) by DCA confirms the classification results, where the 7 vegetation groups were also segregated (Fig. 4). Linear response models were dropped because gradients along the first two axes were longer than 4 SD units (Jongman et al., 1987). The length of gradient represented by axis 1 was > 9 SD, indicating a complete turnover in species composition along this gradient. Therefore, DCA was the appropriate ordination method or indirect gradient analysis to be used. The four DCA axes explained 5.3%, 3.7%, 2.8% and 2.6% of the total variation in the species data, respectively. This low percentage of variance explained by the axes was attributed to the many zero values in the vegetation data set. It can be observed that the eigenvalue for the first DCA axis was high, indicating that it captured the greater proportion of the variation in species composition among stands. It is clear that group C1 occupied the positive end of the first DCA axis, while groups B1 and B2 occupied the negative end. This arrangement may explain a gradient of increasing soil salinity and moisture content (Tab. 4), where stands of group C1 were located along the Red Sea coast transect, while B1 and B2 in the inland desert of Wadi Gimal-Aswan-Wadi Kharit transect. The first DCA axis was positively correlated with electrical conductivity $(r=0.29\dot{7})$, sodium (r=0.342), potassium (r=0.307), calcium (r=0.296), magnesium (r=0.318), chlorides (r= (0.217), moisture contents (r=0.418) and sulfates (r=0.612), and negatively with pH (r=-0.167) and gravels (r=-0.249). The second axis was positively correlated with sulfates (r=0.172) and organic matter (r=0.218).

Comparison between northern and southern parts of Eastern Desert

Tab. 5 displayed the floristic composition between two geographically distant (253 km) parts (northern and southern) of the Eastern Desert. Whereas the southern part represented by the 4 transects included in this study, the northern part (c. 28,800 km²; 30° 05' - 28° 21' N and 31° 20'- 33° 50' E) included three transects; Cairo-Suez (T1N; 112 species), Korimat-Zaafarana (T2N; 111 species) and Sheikh Fadl-Ras Gharib (T3N; 54 species) mainly in the limestone part of this desert (Abdel-Aleem, 2013). Altogether, 60 species were in common, 103 species confined to the northern part, and 34 to the southern part.

Four trees: Acacia tortilis subsp. raddiana, Tamarix aphylla, T. nilotica and Calotropis procera exhibited a wide range of distribution as they recorded in the northern and southern parts. While 9 tree species confined to the southern part, and do not penetrate northwards (e.g., Avicennia marina, Hyphaene thebaica, Balanites aegyptiaca, Moringa peregrina), the northern part devoid of any characteristic tree species. Twentyeight shrubby species were recorded in both areas and included amongst others: Zilla spinosa, Zygophyllum coccineum, Caroxylon imbricatum, Suaeda monoica, Zygophyllum album and Pulicaria incisa. Whereas 33 species were confined to the northern part, 9 species characterized the southern part. Perennial herbs were represented by 19 species (Tab. 5), of which 6 were in common (e.g., Phragmites australis, Citrullus colocynthis, Stipagrostis plumosa), 7 species showed consistency to northern part (e.g., Lavandula stricta, Lasiurus scindicus, Aeluropus lagopoides), and 6 species to the southern part (e.g., Juncus rigidus, Aeluropus littoralis) which inhabited wet and saline habitats. The annual herbs (96 species) constituted the major component of the floristic diversity and structure; 22 were in common, 64 species confined to the northern part and 10 species confined to the southern. The northern part included Conyza bonariensis, Emex spinosa, Phalaris paradoxa, Lolium Cichorium endivia, Amaranthus viridis, perenne, Spergularia marina and Avena fatua which are among the common weeds of the Egyptian arable lands.



Fig. 4. First two axes of the DCA ordination of 142 stands with the 7 vegetation groups (A-D2) separated by cluster analysis superimposed.

Discussion

The classification and ordination analyses proposed that the vegetation of the study area can be divided into 7 major vegetation groups (plant communities). The members of each pair of groups are, in some cases, linked together by having one of the dominant species in common. It can be noted that certain vegetation groups characterized one or more of the studied transects; group (A) in Idfu-Mersa Alam transect (T2), groups (B1), (B2) and (D2) in Aswan-Kharit-Gimal transct (T3), groups (C1) and (C2) in Qusier-Safaga transect along Red Sea coast (T4), and group (D1) was widely distributed in the study area including T1, T2 and T4. It can be noted that the salt-tolerant plant Tamarix nilotica characterized vegetation group (C1) and (C2) form hillocks of huge sizes, and vigorously growing southwards (Springuel et al., 1991) representing the natural climax community type of the desert wadis with deep deposits and an underground water reserve (Kassas and Zahran, 1962). Tamarix has been identified as a major cause of salt accumulation on the soil surface (Springuel and Ali, 1990), and concentrating a high amount of sodium chloride in specialized glands within its leaves (Bosabalidis, 1992). In addition, there is a relationship between the amount of *Tamarix* litter and the electric conductivity of soil (Briggs et al., 1993). Meanwhile, the lower number of recorded species in vegetation group (C1) inhabiting the coastal plains of the Red Sea may be related to its high soil salinity. Such salinity stress on floristic diversity in the study area and related areas was reported by Moustafa and Klopatek (1995) and Shaltout et al. (1997). Most of the identified vegetation groups have very much in common with that recorded in some wadis of the Eastern Desert (Salama et al., 2012, 2013), Western Desert (Abd El-Ghani, 2000; Bornkamm and Kehl, 1990), in south Sinai region (Moustafa and Zaghloul, 1996) and in northwestern Negev, Israel (Tielbörger, 1997).

In extreme deserts, as in the study area, the plant growth is triggered mainly by rain, and thus is as scarce and unpredictable as the precipitation itself. Vegetation develops in 'contracted mode' (Monod, 1954) only in habitats receiving runoff water including wadis, depressions and channels- contracted desert (Shmida, 1985). This highly dynamic vegetation is neither permanent nor seasonal, but is accidental (Bornkamm, 2001; Bullard, 1997; Kassas, 1966). The vegetation structure in the study area is relatively simple, in which the species have to withstand the harsh environmental conditions. This it can be reflected by the presence of several highly adapted, drought-resistant species. The floristic diversity of the study area included 94 species of the vascular plants (67 perennials and 27 annuals) indicating the predominance of perennials. Asteraceae, Fabaceae, Poaceae, Zygophyllaceae and Chenopodiaceae were the species-rich families which formed the major component of the flora. The first three families represent the most common in the Mediterranean North African flora (Quézel, 1978; White, 1993). These findings were in line with those of Salama et al. (2012, 2013) in the Eastern Desert, and Abd El-Ghani and Fahmy (1998) in south Sinai, and Salama *et al.* (2005) along the western Mediterranean coast.

Chorological analysis revealed that the Saharo-(37.2% monoregional, Arabian element 28.7% biregional, 11.7% as pluriregional floras) forms the major component of the floristic structure along the four transects. That is because the study area lies within the Saharo-Arabian region of the Holarctic Kingdom (White, 1993). The results were in agreement with those of El-Demerdash et al. (1990), Fossati et al. (1998) and Salama et al. (2012) who concluded that plants of Saharo-Arabian region constituting the shrub layer as good indicators for desert environmental conditions, while species of Mediterranean origin (either mono, bi or pluriregional) flourish in more mesic conditions.

Comparing the results of floristic diversity in the study area (south of the Eastern Desert) with that in the northern part (Abdel-Aleem, 2013) indicated that 60 species were in common, 103 confined to the northern part, while 34 species were consistent to the southern part. So, the floristic diversity in the northern part is three times higher than that of the southern part of the Eastern Desert, which may be attributed to the mild climatic conditions prevailing in the north. Also, increasing the aridity southwards plays a paramount role in reducing floral diversity. On the other hand, 60% of the northern vegetation (not present in south) was represented as annual herbs. Decreased numbers of annuals in the southern part of the Eastern Desert can be attributed to the environmental aridity and thermal continentality which increases from north to south (Abd El-Ghani, 1998).

Vast areas in the Egyptian deserts (Western, Eastern, and Sinai) were subjected to land reclamation due to increased population growth (Biswas, 1993). In the study area, agricultural processes were practiced in the deltaic parts of several wadis such as Wadi Kherit, W. Natash and W. El-Shikh. As the land reclamation processes entail an almost complete change of the environmental factors, several common weeds of the agro-ecosystem were recorded (e.g., Cynodon dactylon, Malva paviflora, Dicanthiun annulatum, Cyperus rotundus, Sonchus oleraceus and Chenopodium murale). Thus, weeds find the new conditions favorable for their growth. Close to the boundaries of the desert in this study, xerophytic species naturally grow among the weeds of the cultivation. This indicated that these species are native to the natural desert vegetation and can remain after the reclamation process. Therefore, the reclaimed lands found at the desert outskirts can be considered as transitional areas of the succession process between the old cultivated lands and that of the desert (Abd El-Ghani *et al.*, 2013b; Shaheen, 2002).

As for species abundance, Zilla spinosa, Acacia tortilis subsp. raddiana, Morettia philaeana, Zygophyllum coccineum, Caroxylon imbricatum and Citrullus colocynthis (especially in Wadi Natash) had the highest Q-Values (P= 0.61, 0.36, 0.3, 0.23, 0.22, and 0.21%, respectively). This result was in line with that obtained by Abd El-Ghani et al. (2013a) and Salama et al. (2012) in northern and central parts of the Eastern Desert, and Springuel *et al.* (2006) in the south-eastern part of this desert. *Acacia tortilis* subsp. *raddiana, Morettia philaeana* and *Citrullus colocynthis* were completely absent in Red Sea transect, while the presence of the salttolerant species such as *Tamarix nilotica, Limonium* axillare, Arthrocnemum macrostachyum, Juncus rigidus and Nitraria retusa with high presence values in the Red Sea transect indicated its salinized habitat. The record of Avicennia marina dominating the mangal vegetation along the Red Sea coast (T4) is notable, and was documentd by Zahran and Willis (2009).

Tab. 1. Species composition of the 4 transects classified according to the different functional groups, together with their presence values (P%), chorology, Q-values and occurrences. T1= Qena-Safaga transect; T2= Idfu-Marsa Alam transect; T3= Aswan-Kharit-Gimal transect and T4= Red Sea transect. Choro= Chorology (SA= Saharo-Arabian, SZ= Sudano-Zambezian, M= Mediterranean, IT= Irano-Turanian, ES= Euro-Siberian, SU= Sudanian, GC= Gueno-Cungo, COSM= Cosmopolitan, PAN= Pantropical, PAL= Palaeotropical). Occ= Occurrence (D=Dominant, VC=Very common, C=Common, O=Occasional, S=Sporadic)

<u>Crassing</u>	p%	for each	transect	t	Chara	01	0
Species	T1	T2	T 3	T 4	Cnoro	Q-value	Ücc
1	2	3	4	5	6	7	8
Species present in all transects							
Shrubs							
Zilla spinosa (L.) Prantl.	81.8	96.4	73.9	15.2	SA	0.61	D
Zygophyllum coccineum L.	59.1	3.6	8.7	30.4	SA	0.23	D
<i>Caroxylon imbricatum</i> (Forssk.) Akhani & F. H. Roalson	45.5	67.9	2.2	2.2	SA	0.22	D
Lotus hebranicus Hochst. ex Brand	13.6	14.3	17.4	13	М	0.15	VC
Aerva javanica (Burm. F.) Juss ex Schult.	18.2	25	8.7	4.3	SA	0.12	VC
Leptadenia pyrotechnica (Forssk.) Decne.	18.2	7.1	6.5	2.2	SA+SZ	0.07	C
Annual Herbs	10.2	,	0.9	2.2	011102	0.07	U
Astragalus vogelii (Webb.) Bornm.	9.1	21.4	13	6.5	SA	0.12	VC
Tetraena simplex (L.) Beier & Thulin	91	28.6	87	2.2	SA+SZ	0.11	VC
Polycarpaea repens (Forssk.) Asch. &	4.5	3.6	4.3	6.5	SA	0.05	C
Species present in three transects							
Trees							
Acacia tortilis (Forssk.) Havne subsp.							
raddiana (Savi) Brenan	0	46.4	65.2	17.4	SA	0.36	D
Tamarix aphylla (L) H Karst	0	10.7	26.1	174	SA+IT	0.16	VC
T nilotica (Ehreub) Bunge	18.2	0	/ 3	30.4		0.16	VC
Calatratis process (Aiton) W.T. Aiton	4.5	71	4.5	0	54+57	0.04	0
Caloropis procera (Alton) w. 1. Alton	4.)	/.1	0.5	0	SATSL	0.04	0
Surubs	18.2	464	0	2.2	SA	0.13	VC
Pulicaria un dulata (L) C A Mey	0	20.2	10.9	2.2	SA SA	0.12	VC
Deninum tumidum Earol	0	39.5	15.2	2.2	MISA	0.12	ve C
Panuam targuam Forssk.	10.2	5.0	13.2	0./	M+SA	0.08	C
Dentradenus baccatus Denie	18.2	10.7	2.2	6.5	SA	0.06	C
Pergularia tomentosa L.	13.6	10.7	2.2	0	SA	0.05	C
Suaeda monoica Forssk. ex J. F. Gmel.	0	7.1	2.2	6.5	SA+SZ	0.04	0
Cleome droserifolia (Forssk.) Delile	4.5	0	4.3	6.5	SA+IT	0.04	0
Perennial Herbs				-			~
Citrullus colocynthis (L.) Schrad.	22.7	35.7	32.6	0	M+SA+IT	0.21	D
Phragmites australis (Cav.) Trin. ex Steud.	18.2	3.6	0	13	PAN	0.08	С
Monsonia heliotropioides (Cav.) Boiss.	0	3.6	10.9	2.2	SA	0.05	С
Annual Herbs							_
Morettia philaeana (Delile) DC.	31.8	60.7	39.1	0	SA	0.3	D
Schouwia purpurea (Forssk.) Schweinf.	9.1	35.7	8.7	0	SA	0.11	VC
Trichodesma africanum (L.) R. Br.	22.7	25	4.3	0	SA+SZ	0.1	VC
<i>Cotula cinerea</i> Delile	9.1	3.6	19.6	0	IT	0.09	С
Forsskaolea tenacissima L.	18.2	3.6	6.5	0	SA+SZ	0.06	С
Eremobium aegyptiacum (Spreng.) Asch. & Schweinf. ex Boiss.	4.5	10.7	2.2	0	SA	0.04	0
Species present in two transects							
Trees							
Phoenix dactylifera L.	22.7	0	0	2.2	SA+SZ	0.04	0

1	2	3	4	5	6	7	8
Shrubs							
<i>Farsetia stylosa</i> R. Br.	0	10.7	17.4	0	SA+SZ	0.08	С
Pulicaria incisa (Lam.) DC.	27.3	0	4.3	0	SA	0.06	С
Senna italica Mill	0	3.6	17.4	0	SA+SZ	0.06	С
Convolvulus hystrix Vahl	0	0	2.2	8.7	SA	0.04	0
Fagonia indica Burm. F.	0	0	6.5	2.2	SA	0.03	0
Heliotropium bacciferum Forssk.	0	7.1	0	2.2	SA	0.02	О
Perennial Herbs							
Cynodon dactylon (L.) Pers.	13.6	0	0	4.3	COSM	0.04	О
Annual Herbs							
Launaea nudicaulis (L.) Hook. F.	0	0	13	8.7	SA+IT	0.07	С
Asphodelus tenuifolius Cav.	0	7.1	15.2	0	M+SA+IT	0.06	С
Tribulus pentandrus Forssk.	0	14.3	10.9	0	SA+SZ	0.06	С
Malva parviflora L.	0	0	2.2	10.9	M+ES+IT	0.04	О
Cleome amblyocarpa Barratte &Murb.	0	3.6	10.9	0	SA+SZ	0.04	0
Arnebia hispidissima (Lehm.) DC.	4.5	0	0	4.3	SA	0.02	О
<i>Euphorbia granulata</i> Forssk.	0	3.6	4.3	0	M+SA+IT	0.02	0
Reseda pruinosa Delile	0	0	2.2	2.2	SA	0.01	О
Cistanche phelypaea (L.) Cout.	4.5	3.6	0	0	M+SA+IT	0.01	0
Tribulus megistopterus Kralik	4.5	0	2.2	0	SA+SZ	0.01	О
Species present in one transect							
Trees							
Balanites aegyptiaca (L.) Delile	0	0	28.3	0	SA+SZ	0.09	С
Ziziphus spina-christi (L.) Desf.	18.2	0	0	0	SA	0.03	О
Avicennia marina (Forssk.) Vierh.	0	0	0	8.7	SA	0.03	О
Ricinus communis L.	9.1	0	0	0	PAN	0.01	О
Acacia nilotica (L.) Delile	4.5	0	0	0	SU	0.007	S
Capparis decidua (Forssk.) Edgew.	0	0	2.2	0	SA+SZ	0.007	S
Hyphaene thebaica (L.) Mart.	0	0	0	2.2	SA+SZ	0.007	S
Moringa peregrina (Forssk.) Fiori	4.5	0	0	0	SZ+GC	0.007	S
Shrubs							
Zygophyllum album L.	0	0	0	26.1	M+SA+IT	0.08	С
Nitraria retusa (Forssk.) Asch.	0	0	0	26.1	SA	0.08	С
Limonium axillare (Forssk.) Kuntze	0	0	0	21.7	SA+SZ	0.06	С
Crotalaria aegyptiaca Benth.	0	0	0	10.9	SZ	0.04	0
Arthrocnemum macrostachyum (Moric.) K. Koch	0	0	0	13	M+SA+IT	0.04	Ο
Salvadora persica L.	0	0	8.7	0	SA+SZ	0.03	0
Cornulaca monacantha Delile	0	0	0	6.5	SA	0.02	О
Artemisia judaica L.	9.1	0	0	0	SA	0.01	0
Senna holosericea (Freseu) Greuter	0	0	4.3	0	SA	0.01	0
Fagonia mollis Delile	9.1	0	0	0	SA	0.01	0
Iphiona mucronata (Forssk.) Asch. & Schweinf.	0	7.1	0	0	SA	0.01	О
Atriplex leucoclada Boiss.	4.5	0	0	0	SA+IT	0.007	S
Fagonia bruguieri DC.	4.5	0	0	0	SA+IT	0.007	S
Capparis spinosa L.	0	0	0	2.2	M+SA+SZ	0.007	S
Chrozophora oblongifolia (Delile) Spreng.	4.5	0	0	0	M+SA	0.007	S
Oxystelma esculentum (L.F.) R. Br.	0	0	2.2	0	SZ+GC	0.007	S
Caroxylon villosum (Schult.) Akhani & E. H. Roskon	4.5	0	0	0	M+SA+IT	0.007	S
Taverniera aegyptiaca Boiss.	0	0	0	2.2	SA	0.007	S

1	2	3	4	5	6	7	8
Perennial Herbs							
Aeluropus littoralis (Gouan) Parl.	0	0	0	6.5	M+IT	0.02	0
Imperata cylindrica (L.) Raeusch	9.1	0	0	0	PAN	0.01	0
Juncus rigidus Desf.	0	0	0	2.2	M+SA+IT	0.007	S
Leptochloa fusca (L.) Kunth	0	0	0	2.2	PAL	0.007	S
<i>Stipagrostis plumosa</i> (L.) Munro ex T. Anderson	4.5	0	0	0	M+SA+IT	0.007	S
Dichanthum annulatum (Forssk.) Stapf	4.5	0	0	0	PAL	0.007	S
Cyperus rotundus L.	0	0	0	2.2	PAN	0.007	S
Typha domingensis (Pers.) Poir. ex Steud.	4.5	0	0	0	PAN	0.007	S
Annual Herbs							
Astragalus eremophilus Boiss.	0	0	30.4	0	SA	0.1	С
Hippocrepis constricta Knuze	0	0	6.5	0	M+SA+IT	0.02	0
Lupinus digitatus Forssk.	0	0	6.5	0	М	0.02	0
<i>Polycarpaea robbairea</i> (Kuntze) Greuter & Burdet	0	0	6.5	0	SA	0.02	О
Launaea amal-aminae N. Kilian	0	0	4.3	0	SA	0.01	0
Echium horridum Batt.	4.5	0	0	0	SA	0.007	S
Chenopodium album L.	4.5	0	0	0	COSM	0.007	S
Ch. murale L.	0	0	0	2.2	COSM	0.007	S
Launaea capitata (Spreng.) Dandy	0	0	2.2	0	SA	0.007	S
Filago desertorum Pomel	4.5	0	0	0	SA+IT	0.007	S
Glinus lotoides L.	0	0	2.2	0	PAL	0.007	S
<i>Oligomeris linifolia</i> (Vahl.) ex Hornew J. F. Macbr.	0	0	0	2.2	SA+SZ	0.007	S
Sonchus oleraceus L.	0	0	0	2.2	COSM	0.007	S

Tab. 2. Species composition of the obtained 7 vegetation groups, together with their presence values (p%). Figures in bold are the dominant species that have the highest p%.

Vegetation groups	Α	B1	B2	C1	C2	D1	D2
Total number of stands	18	18	7	41	8	31	19
Total number of species	41	26	30	32	19	53	20
1	2	3	4	5	6	7	8
Species present in 6 groups							
Zilla spinosa	100	94.4	100		12.5	90.3	78.9
Acacia tortilis subsp. raddiana	38.9	77.8	71.4	19.5		16.1	63.2
Species present in 5 groups							
Tamarix aphylla	16.7			14.6	12.5	3.2	63.2
Zygophyllum coccineum	5.6			7.3	100	51.6	21.1
Aerva javanica	27.8		14.3	12.2		16.1	5.3
Pulicaria undulata	27.8	22.2		4.9		9.7	15.8
Lotus hebranicus	27.8	5.6	57.1	7.3		25.8	
Species present in 4 groups							
Citrullus colocynthis	44.4	83.3	57.1			9.7	
Morettia philaeana	94.4	83.3	85.7			12.9	
Pergularia tomentosa	5.6	5.6	14.3			12.9	
Tetraena simplex	50	11.1	14.3			9.7	
Tribulus pentandrus	27.8	11.1	14.3				5.3
Caroxylon imbricatum	83.3	22.2		2.4		35.5	
Astragalus vogelii	38.9		85.7			9.7	5.3
Forsskaolea tenacissima	11.1		28.6			9.7	5.3
Phragmites australis	11.1			7.3	37.5	9.7	
Tamarix nilotica	5.6			22	100	6.5	

50

1	2	3	4	5	6	7	8
Malva parviflora	5.6			2.4		9.7	5.3
Leptadenia pyrotechnica		5.6		4.9		12.9	15.8
Panicum turgidum			14.3	12.2		12.9	10.5
Species present in 3 groups							
Asphodelus tenuifolius	5.6	22.2	57.1				
Astragalus eremophilus	11.1	33.3	85.7				
Cotula cinerea	16.7	16.7	85.7				
Monsonia heliotropioides	11.1		57.1			3.2	
Pulicaria incise	11.1		28.6			12.9	
Schouwia purpurea	61.1		42.9			6.5	
Trichodesma africanum	44.4		14.3			16.1	
Eremohium aegyptiacum	16.7		14.3				5.3
Farsetia stylosa	167		85.7				10.5
Arnehia histoidissima	5.6		0.7	24		32	10.9
Fagonia indica	5.6			2.1 49		3.2	
Comodore dactulore	5.6			ч.)	25	5.2	
Example the holes	72.2				12.5	12.0	
	/ 2.2	= (1/2	2 /	12.5	12.9	
Polycarpaea robbairea		5.6	14.3	2.4		0.7	
Convolvulus hystrix		5.6		2.4		9.7	
Calotropis procera		11.1				3.2	15.8
Launaea nudicaulis			85.7	2.4		9.7	
Polycarpaea repens			28.6	2.4		12.9	
Nitraria retusa				17.1	50		5.3
Ochradenus baccatus					12.5	19.4	5.3
Species present in 2 groups							
Euphorbia granulata	11.1	5.6					
Suaeda monoica	16.7			7.3			
Cistanche phelypaea	5.6					3.2	
Ricinus communis	5.6					3.2	
Cleome amblyocarpa		11.1	57.1				
Hippocrepis constricta		5.6	28.6				
Lupinus digitatus		5.6	28.6				
Senna italica		44.4	14.3				
Artemisia judaica		5.6				3.2	
Fagonia mollis		5.6				3.2	
Limonium axillare		,		19.5	25	2.1	
Zvgothvllum alhum				24.4	25		
Crotalaria aegyptiaca				73		65	
Heliotropium hacciferum				24		6.5	
Reseda pruinosa				2.1		2.2	
Rescui prumosa Relamitas gometriasa				2.4		5.2	(2.2
				2.4	25	12.0	65.2
Cleome aroserijolia					25	12.9	
Phoenix dactylijera					12.5	16.1	
Species present in one group	5 (
Echium horriaum	5.6						
Glinus lotoides	5.6						
Oxystelma esculentum	5.6						
Caroxylon villosum	5.6						
Stipagrostis plumosa	5.6						
Tribulus megistopterus	11.1						
Chenopodium album		5.6					
Filago desertorum		5.6					

1	2	3	4	5	6	7	8
Launaea capitata			14.3				
L. amal-aminae			28.6				
Aeluropus littoralis				7.3			
Arthrocnemum macrostachyum				14.6			
Avicennia marina				9.8			
Capparis spinosa				2.4			
Cornulaca monacantha				7.3			
Senna holosericea				4.9			
Chenopodium murale					12.5		
Cyperus rotundus					12.5		
Hyphaene thebaica					12.5		
Juncus rigidus					12.5		
Leptochloa fusca					12.5		
Sonchus oleraceus					12.5		
Acacia nilotica						3.2	
Atriplex leucoclada						3.2	
Chrozophora oblongifolia						3.2	
Dichanthum annulatum						3.2	
Fagonia bruguieri						3.2	
Imperata cylindrica						6.5	
Iphiona mucronata						6.5	
Moringa peregrina						3.2	
Oligomeris linifolia						3.2	
Taverniera aegyptiaca						3.2	
Typha domingensis						3.2	
Ziziphus spina-christi						12.9	
Capparis decidua							5.3
Salvadora persica							21.1

Tab. 3. Mean values, standard deviations (\pm SD) and ANOVA values of the soil variables in the vegetation groups (A-D2) of the study area. EC=Electrical conductivity (mS.cm⁻¹), soil fractions (%), CS=Coarse sand, FS=Fine sand, OM=Organic matter, Na⁺, K⁺, Ca⁺², Mg⁺², Cl⁻ and HCO₃⁻¹ (mg.g⁻¹ d.wt. soil), SO₄⁻² (µg.g-1 d.wt. soil), OM (%), *= p<0.05, **= p<0.01.

Soil	Total			,	Vegetation group	DS			r 1
Factors	Means	Α	B1	B2	CI	C2	D1	D2	F value
Gravel	11.27±10.62	13.03±10.42	12.03±8.62	6.91±3.58	9.56±10.07	7.44±14.14	16.22±11.67	7.76±9.86	2.223*
CS	20.97±16.48	15.37±10.64	21.67±11.86	20.49±9.31	25.44±21.94	8.53±7.56	24.06 ± 14.88	16.36 ± 14.60	2.145
FS	9.31±6.19	5.59±2.92	11.40±7.89	12.83±4.92	9.70±6.45	7.58±6.33	8.39±5.41	10.99±6.26	2.421**
Silt	35.52±16.51	39.59±13.71	34.16±13.09	44.85±9.03	33.44±17.59	48.35±14.83	26.38±12.96	43.52±19.20	4.36**
Clay	22.91±18.13	26.43±13.68	20.74±19.99	14.93±6.06	21.86±19.35	28.10±14.60	24.95±23.67	21.37±9.70	0.592
WC	2.61 ± 4.40	1.20 ± 1.14	1.51 ± 1.24	1.27 ± 0.80	5.16±6.07	5.33±8.71	1.58 ± 2.14	0.53 ± 1.08	5.186**
OM	1.09 ± 0.10	1.13±0.10	1.12±0.04	1.01 ± 0.10	1.08 ± 0.12	1.12 ± 0.03	1.11 ± 0.09	1.03 ± 0.09	3.639**
pН	7.80 ± 0.48	8.02±0.34	7.97±0.68	7.67±0.81	7.78±0.45	7.20±0.48	7.77±0.38	7.86±0.16	3.599**
EC	1.22±3.35	0.32±0.11	0.22 ± 0.05	0.21±0.05	1.62±2.27	8.78±10.77	0.60 ± 1.00	0.39±0.51	11.296**
Na ⁺	0.83±2.33	0.11±0.07	0.05 ± 0.02	0.05 ± 0.02	1.29 ± 1.73	5.86±7.35	0.31±0.66	0.25 ± 0.48	10.916**
K^+	0.08±0.13	0.04 ± 0.01	0.03 ± 0.01	0.02 ± 0.00	0.11±0.10	0.21±0.27	0.05±0.03	0.11±0.24	3.606**
Ca ⁺²	0.63±1.51	0.31±0.32	0.16 ± 0.04	0.15 ± 0.05	0.83±0.94	3.80 ± 4.98	0.38±0.67	0.17 ± 0.14	9.575**
Mg ⁺²	0.26±0.55	0.08±0.03	0.10 ± 0.04	0.14 ± 0.05	0.43±0.71	1.04 ± 1.38	0.16±0.17	0.05±0.03	5.53**
Cl	1.41 ± 6.04	0.09±0.07	0.10 ± 0.03	0.10 ± 0.03	1.51 ± 4.12	15.08±20.02	0.33 ± 0.81	0.15±0.18	10.51**
HCO ₃ ⁻	0.18±0.06	0.18 ± 0.05	0.16 ± 0.05	0.16±0.08	0.18±0.06	0.20±0.12	0.17±0.05	0.18 ± 0.04	0.64
SO4-2	5.47±7.36	1.36±1.24	1.44±1.66	0.67±0.96	10.27±8.27	15.11±10.12	4.29±6.64	2.46±2.68	12.009**

53

Tab. 4. Simple linear correlation coefficient (r) between the soil variables and DCA axes. *= p<0.05, **: p<0.01. For soil factors abbreviations and units, see Tab. 4.

Soil variables	DCA axis 1	DCA axis 2
pН	-0.167*	-0.15
EC	0.297**	0.116
Na ⁺	0.342**	0.146
K^+	0.307**	0.026
Ca ⁺²	0.296**	0.119
Mg ⁺²	0.318**	-0.015
Cl	0.217**	0.098
HCO ₃ -	0.119	0.072
SO4 ⁻²	0.612**	0.172^{*}
MC	0.418**	-0.058
OM	-0.024	0.218**
Gravels	-0.249**	0.022
CS	0.034	-0.111
FS	-0.013	0.081
Silt	0.073	0.009
Clay	0.053	0.052

Tab. 5. Floristic diversity between the northern and southern parts of the Eastern Desert. (¹ =Data from Abdel Aleem, 2013); T1N= Cairo-Suez transect; T2N= Korimat-Zaafarana transect, T3N= Sheikh Fadl-Ras Gharib transect; T1S= Qena-Safaga transect; T2S= Idfu-Marsa Alam transect; T3S= Aswan-Kharit-Gimal transect and T4S= Red Sea transect.

c · ·		North					
Species	T1N	T2N	T3N	T1S	T2S	T3S	T4S
1	2	3	4	5	6	7	8
Species present in Both Areas							
Acacia tortilis (Forssk.) Hayne subsp. raddiana Savi (Brenan)	35	18.2	16.7	0	46.4	65.2	17.4
<i>Tamarix aphylla</i> (L.) H Karst.	0	0	16.7	0	10.7	26.1	17.4
Tamarix nilotica (Ehrenb.) Bunge	40	59.1	30	18.2	0	4.3	30.4
Calotropis procera (Aiton) W. T. Aiton	30	0	0	4.5	7.1	6.5	0
Zilla spinosa (L.) Prantl.	95	81.8	83.3	81.8	96.4	73.9	15.2
Zygophyllum coccineum L.	85	63.6	83.3	59.1	3.6	8.7	30.4
Caroxylon imbricatum (Forssk.) Akhani & E. H. Roalson	0	22.7	40	45.5	67.9	2.2	2.2
Aerva javanica (Burm. F.) Juss. ex Schult.	10	0	10	18.2	25	8.7	4.3
Lotus hebranicus Hochst. ex Brand	0	0	6.7	13.6	14.3	17.4	13
Leptadenia pyrotechnica (Forssk.) Decne.	0	0	10	18.2	7.1	6.5	2.2
Pulicaria undulata (L.) C. A. Mey	50	77.3	40	0	39.3	10.9	2.2
Panicum turgidum Forssk.	55	22.7	0	0	3.6	15.2	8.7
Suaeda monoica Forssk. ex J. F. Gmel.	0	13.6	0	0	7.1	2.2	6.5
Ochradenus baccatus Delile	55	59.1	56.7	18.2	0	2.2	6.5
Cleome droserifolia (Forssk.) Delile	0	0	3.3	4.5	0	4.3	6.5
<i>Fagonia indica</i> Burm. F.	20	22.7	10	0	0	6.5	2.2
Heliotropium bacciferum Forssk.	30	4.5	6.7	0	7.1	0	2.2
Zygophyllum album L.	0	9.1	0	0	0	0	26.1
Nitraria retusa (Forssk.) Asch.	0	4.5	0	0	0	0	26.1
Capparis spinosa L.	0	4.5	0	0	0	0	2.2
Taverniera aegyptiaca Boiss.	0	9.1	0	0	0	0	2.2
Crotalaria aegyptiaca Benth.	20	0	0	0	0	0	10.9
Cornulaca monacantha Delile	5	0	0	0	0	0	6.5
Pergularia tomentosa L.	25	40.9	6.7	13.6	10.7	2.2	0
Senna italica Mill.	15	0	0	0	3.6	17.4	0
Pulicaria incisa (Lam.) DC.	10	18.2	0	27.3	0	4.3	0
Iphiona mucronata (Forssk.) Asch. & Schweinf.	20	31.8	0	0	7.1	0	0
Artemisia judaica L.	5	9.1	10	9.1	0	0	0

			,	_	,	_	0
1 Tarris and the Dality	2	3	4	5	6	7	8
Fagonia mouis Deme	20	50 26 /i	15.5	9.1	0	0	0
Atripler leucoclada Boise	0	0	45.5	4.5	0	0	0
Chrosothora chloraifolia (Dolilo) Sprong	10	0	0.7	4.5	0	0	0
Dhugamites australis (Cox) Trip. or Stoud	10	12.6	2.2	18.2	2.6	0	12
Caracter dactular (L) Dara	10	0	0	10.2	<u> </u>	0	13
Leptochlog fuese (L.) Kunth	10	0	0	15.0	0	0	4.5
Citrullus colorunthis (L.) Schrod	5	9.1 27.2	167	22.7	25.7	22.6	2.2
Stip agreetic plumeca (L.) Mupro av T. Anderson)	4.5	2.2	/ 5	55.7	52.6	0
Imperata culindrica (L.) Resusch	0	4.5	0	4.) 9.1	0	0	0
Tatuana cimplex (L.) Roior & Thulin	60	19.0	20	9.1	28.6	87	22
Actuageduc upgelij (Wohh) Bornm	00	0.1	10	9.1	20.0	12	6.5
Launaea nudicaulis (L) Hook E	25	50.1	67	9.1	0	13	87
Pacada truinaca Dolilo	2)	39.1 4.5	10	0	0	15	0.7
Malua transform	ر 15	4.5	10	0	0	2.2	10.0
Maiva parvijiora L.	15	65.5	0	0	0	2.2	10.9
Chamata dium munda L	10	45.5	0	0	0	0	2.2
Chenopolium murale L.	10	21.0	12.2	0	2(10 (2.2
	10	51.8	15.5	9.1	5.0 2.6	19.6	0
Trishedown - frierwym (L.) P. Pr	25	4.5	20	10.2	5.0 25	6.5	0
Irrenodesma ajricanum (L.) K. Br.	25	40.9	20	22.7	25	4.5	0
<i>Eremobium aegyptacum</i> (Spreng.) Ascn. & Schweinr. ex Boiss.	0	4.5	0	4.5	10.7	2.2	0
<i>Cleome amblyocarpa</i> Barratte & Murb.	25	22.7	6.7	0	3.6	10.9	0
Euphorbia granulata Forssk.	0	9.1	0	0	3.6	4.3	0
Asphodelus tenuifolius Cav.	20	0	0	0	7.1	15.2	0
<i>Tribulus megistopterus</i> Kralik	0	4.5	0	4.5	0	2.2	0
Glinus lotoides L.	0	40.9	6.7	0	0	2.2	0
<i>Polycarpaea robbairea</i> (Kuntze) Greuter & Burdet	0	0	6.7	0	0	6.5	0
Hippocrepis constricta Knuze	0	4.5	0	0	0	6.5	0
<i>Launaea capitata</i> (Spreng.) Dandy	0	36.4	0	0	0	2.2	0
<i>Cistanche phelypaea</i> (L.) Cout.	15	31.8	6.7	4.5	3.6	0	0
Filago desertorum Pomel	0	9.1	0	4.5	0	0	0
Chenopodium album L.	5	0	0	4.5	0	0	0
Species present in Northern Area							
Atriplex halimus L.	15	40.9	36.7	0	0	0	0
Anabasis setifera Moq.	40	31.8	33.3	0	0	0	0
Farsetia aegyptia Turra	60	50	13.3	0	0	0	0
Haloxylon salicornicum (Moq.) Bung ex Boiss.	65	59.1	56.7	0	0	0	0
Launaea spinosa (Forssk.) Sch. Bip. ex Kuntze	10	18.2	3.3	0	0	0	0
Retama raetam (Forssk.) Webb & Berthel.	45	27.3	13.3	0	0	0	0
Hyoscyamus muticus L.	50	22.7	3.3	0	0	0	0
Senecio glaucus L.	25	36.4	3.3	0	0	0	0
Astragalus trigonus DC.	0	9.1	3.3	0	0	0	0
Calligonum polygonoides L.	0	9.1	26.7	0	0	0	0
Deverra tortuosa (Desf.) DC.	35	0	3.3	0	0	0	0
Hyoscyamus boveanus (Dunal) Asch. & Schweinf.	0	0	3.3	0	0	0	0
Achillea fragrantissima (Forssk.) Sch. Bip.	5	13.6	0	0	0	0	0
	40	9.1	0	0	0	0	0
Anabasis articulata (Forssk.) Moq.			0	0	0	0	0
Anabasis articulata (Forssk.) Moq. Cynanchum acutum L.	15	9.1	0	0	0	0	U
Anabasis articulata (Forssk.) Moq. Cynanchum acutum L. Diplotaxis acris (Forssk.) Boiss.	15 10	9.1 4.5	0	0	0	0	0
Anabasis articulata (Forssk.) Moq. Cynanchum acutum L. Diplotaxis acris (Forssk.) Boiss. Ephedra alata Decne.	15 10 10	9.1 4.5 9.1	0 0 0	0 0 0	0 0 0	0 0 0	0
Anabasis articulata (Forssk.) Moq. Cynanchum acutum L. Diplotaxis acris (Forssk.) Boiss. Ephedra alata Decne. Fagonia arabica L.	15 10 10 10	9.1 4.5 9.1 4.5	0 0 0	0 0 0	0 0 0	0 0 0	0 0 0
Anabasis articulata (Forssk.) Moq. Cynanchum acutum L. Diplotaxis acris (Forssk.) Boiss. Ephedra alata Decne. Fagonia arabica L. Gymnocarpos decandrus Forssk.	15 10 10 10 5	9.1 4.5 9.1 4.5 9.1	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0
Anabasis articulata (Forssk.) Moq. Cynanchum acutum L. Diplotaxis acris (Forssk.) Boiss. Ephedra alata Decne. Fagonia arabica L. Gymnocarpos decandrus Forssk. Haplophyllum tuberculatum (Forssk.) A. Juss.	15 10 10 10 5 10	9.1 4.5 9.1 4.5 9.1 9.1	0 0 0 0 0	0 0 0 0 0	0 0 0 0 0	0 0 0 0 0	0 0 0 0 0

1	2	3	4	5	6	7	8
Nauplius graveolens (Forssk.) Wiklund	10	31.8	0	0	0	0	0
Rumex vesicarius L.	30	22.7	0	0	0	0	0
<i>Reichardia tingitana</i> (L.) Roth	25	18.2	0	0	0	0	0
Agathophora alopecuroides (Delile) Fenzl ex Bunge	0	13.6	0	0	0	0	0
Astragalus sieberi DC.	0	9.1	0	0	0	0	0
Cullen plicata (Delile) C. H Stirt.	0	4.5	0	0	0	0	0
Helianthemum kahiricum Delile	0	4.5	0	0	0	0	0
<i>Reaumuria hirtella</i> Jaub. et Spach	0	31.8	0	0	0	0	0
Ephedra aphylla Forssk.	10	0	0	0	0	0	0
Artemisia monosperma Delile	20	0	0	0	0	0	0
Convolvulus lanatus Vahl	5	0	0	0	0	0	0
Helianthemum lipii (L.) Dum-Cours.	5	0	0	0	0	0	0
Kickxia aegyptiaca (Dum.) Nabelek	45	0	0	0	0	0	0
Phagnalon barbeyanum Asch. & Schweinf.	5	0	0	0	0	0	0
Pluchea dioscoridis (L.) DC.	15	0	0	0	0	0	0
Zygophyllum decumbens Delile	10	0	0	0	0	0	0
Lavandula coronopifolia Delile	5	4.5	0	0	0	0	0
Erodium glaucophyllum (L.) L'Hér. Poirin	20	4.5	0	0	0	0	0
Erodium oxyrrhynchum M. Bieb. Lam.	10	18.2	0	0	0	0	0
Lasiurus scindicus Henrad	5	18.2	0	0	0	0	0
Launaea mucronata (Forssk.) Muschl	40	31.8	0	0	0	0	0
Aeluropus lagopoides (L.) Trip, ex Thwaites	0	45	0	0	0	0	0
Pennisetum divisum (Forssk, ex L E Gmel.) Henrard	10	0	0	0	0	0	0
Anastatica hierochuntica L	10	40.9	167	0	0	0	0
Rassia muricata (L.) Asch	30	18.2	33	0	0	0	0
Trigonella stellata Forssk	35	54.5	3.3	0	0	0	0
Centairea aeroptiaca	5	13.6	10	0	0	0	0
Futborbia retusa Forssk	35	22.7	13.3	0	0	0	0
Bassia indica (Wight) A L Scott	15	0	13.3	0	0	0	0
Latus alinoides Delile	15	0	2.2	0	0	0	0
Lotonomic platneart a (Viv.) Die Serm	0	0	2.2	0	0	0	0
Sugada alticizina (U.) Pil. Serii.	0	0	2.2	0	0	0	0
Altherer le deviei i I	5	27.2	5.5	0	0	0	0
Anchusa histoida Eoreola	20	18.2	0	0	0	0	0
Diplotavic hama (Forsole) Boise	20 //5	26 /	0	0	0	0	0
Contained calcitrate I	4)	21.9	0	0	0	0	0
Centaurea cautirapa L.	40	51.0	0	0	0	0	0
Astragatus bombychus Boiss.	10	4.5	0	0	0	0	0
Echineste et inecieire Turre	70	4.)	0	0	0	0	0
Econops spinosissimus Turra	5	4.5	0	0	0	0	0
Emex spinosa (L.) Campd.	5	4.5	0	0	0	0	0
<i>Eroarum matacoiaes</i> (L.) L Her.) 25	9.1	0	0	0	0	0
Matthiola longipetala (Delle) DC.	35	4.5	0	0	0	0	0
Paronychia arabica (L.) DC.	10	18.2	0	0	0	0	0
Plantago ovata Forssk.	>	2/.3	0	0	0	0	0
Pteranthus aichotomus Forssk.	10	18.2	0	0	0	0	0
Convolvulus puoselufolius Desr.	0	4.5	0	0	0	0	0
Astragalus hamosus L.	0	4.5	0	0	0	0	0
Astragalus schimperi Boiss.	0	4.5	0	0	0	0	0
Atractylis mernephtae Asch	0	9.1	0	0	0	0	0
Avena sterilis L.	0	4.5	0	0	0	0	0
Caylusea hexagyna (Forssk.) M.L. Green	0	4.5	0	0	0	0	0
Crypsis aculeata (DC.) Boiss	0	4.5	0	0	0	0	0
Herniaria hemistemon J. Gay	0	4.5	0	0	0	0	0
Ifloga spicata (Forssk.) SchBip.	0	22.7	0	0	0	0	0
Lappula spinocarpos (Forssk.) Asch. ex Kuntze	0	13.6	0	0	0	0	0

1	2	3	4	5	6	7	8
Morettia philaeana (Delile) DC.	0	0	0	31.8	60.7	39.1	0
Schouwia purpurea (Forssk.) Schweinf.	0	0	0	9.1	35.7	8.7	0
Tribulus pentandrus Forssk.	0	0	0	0	14.3	10.9	0
Astragalus eremophilus Boiss.	0	0	0	0	0	30.4	0
Lupinus digitatus Forssk.	0	0	0	0	0	6.5	0
Launaea amal-aminae N. Kilian	0	0	0	0	0	4.3	0
Echium horridum Batt.	0	0	0	4.5	0	0	0

References

- Abd El-Ghani MM (1998). Environmental correlates of species distribution in arid desert ecosystems of eastern Egypt. Journal of Arid Environment 38:297-313.
- Abd El-Ghani MM (2000). Floristics and environmental relations in two extreme desert zones of western Egypt. Global Ecology and Biogeography 9:499-516.
- Abd El-Ghani MM, Amer WM (2003). Soil-vegetation relation-ships in a coastal desert plain of southern Sinai, Egypt. Journal of Arid Environment 55(4):607-28.
- Abd El-Ghani MM, Fahmy AG (1998). Composition of and changes in the spontaneous flora of Feiran Oasis, S. Sinai, Egypt, in the last 60 years. Willdenowia 28:123-134.
- Abd El-Ghani MM, Salama F, Salem B, El-Hadidy A, Abdel-Aleem M (2013a)._Biogeographical relations of a hyperarid desert flora in eastern Egypt. African Journal of Ecology (doi/10.1111/aje.12102).
- Abd El-Ghani MM, Soliman AT, Hamdy RS, Bennoba IH (2013b). Weed flora in the reclaimed lands along the northern sector of the Nile Valley in Egypt. Turkish Journal of Botany 37:464-488.
- Abdel-Aleem M (2013). Floristic analysis and phytogeographical affinities of the Eastern Desert of Egypt. Faculty of Science, Cairo University, Ph D. Diss.
- Abu Al-Izz MS (1971). Landforms of Egypt, The American University in Cairo Press, Cairo, 281 p.
- Allen MM, Stainer ST (1974). Chemical analysis of Ecological Materials, Blackwell Scientific Publications, London, 565 p.
- Al-Sherif EA, Ayesh AM, Rawi SM (2013). Floristic composition, life form and chorology of plant life at Khulais region, Western Saudi Arabia. Pakistan Journal of Botany 45(1):29-38.
- Austin MP (1990). Community theory and competition in vegetation, p. 215-238. In: Grace JB, Tilman D (Eds.). Perspectives on Plant Competition. Academic Press, San Diego.
- Bardsley CE, Lancaster JD (1965). Sulfur, p. 1102-1116. In: Black CA, Evans DD, White JL, Jnsminger LE, Clark FE (Eds.). Methods of soil analysis. Part 2. Agronomy. Series No. 9. Madison, Wisconsin: American Society of Agronomy, Inc.
- Batalha MA, Martins FR (2002). Life-form spectra of Brazilian cerrado sites. Flora 197:452-460.
- Biswas AK (1993). Land resources for sustainable agricultural

development in Egypt. Ambio 22:556-560.

- Black CA (1979). Methods of soil analysis. American Society of Agronomy 2:771-1572.
- Bornkamm R (2001). Allochthonous ecosystems ecosystems without producers. Bocconea 13:201-208.
- Bornkamm R, Kehl H (1990). The plant communities of the Western desert of Egypt. Phytocoenologia 19(2):149-231.
- Bosabalidis AM (1992). A morphological approach to the question of salt gland lifetime in leaves of *Tamarix aphylla* L. Israel Journal of Botany 41:115-121.
- Boulos L (1995). Flora of Egypt, Check list, Al Hadara Publishing, Cairo, 283 p.
- Boulos L (1999). Flora of Egypt, Vol. 1 Azollaceae-Oxalidaceae, Al Hadara Publishing, Cairo, 419 p.
- Boulos L (2000). Flora of Egypt, Vol. 2 Geraniaceae-Boraginaceae. Al Hadara Publishing, Cairo, Egypt, 293p.
- Boulos L (2002). Flora of Egypt, Vol. 3 Verbenaceae-Compositae, Al Hadara Publishing, Cairo, 373 p.
- Boulos L (2005): Flora of Egypt, Vol. 4 Monocotyledons (Alismataceae-Orchidaceae), Al Hadara Publishing, Cairo, 617 p.
- Briggs J, Dickinson G, Murphy K, Pulford I, Belal A, Moalla S, Springuel I, Ghabbour S, Mekki AM (1993). Sustainable development and resource management in marginal environments: natural resources and their use in Wadi Allaqi region of Egypt. Applied Geography 13:259-284.
- Bullard JE (1997). Vegetation and dryland morphology, p. 109-131. In: Thomas DSG (Eds.). Arid Zone Geomorphology. J. Wiley and Sons, Chichester.
- Da Costa RC, De Araújo FS, Lima-Verde LW (2007). Flora and life-form spectrum in an area of deciduous thorn woodland (caatinga) in northeastern. Journal of Arid Environment 68:237-247.
- Dale G (1998). Forest plant diversity at local and landscape scales in the Cascade Mountains of Southwestern Washington. Forest Ecology and Management 109:323-341.
- Danin A, Shmida A, Liston A (1985). Contribution to the flora of Sinia, III. Checklist of the species collected and recorded by the Jerusalem team 1967-1982. Willdenowia 15(1):255-322.
- El-Demerdash MA, Zahran MA, Serag MS (1990). On the ecology of the deltaic Mediterranean coastal land, Egypt. III. The habitat of salt marshes of Damietta-Port Said coastal region. Arab Gulf Journal of Scientific Research

8(3):103-119.

- El Hadidi MN, Fayed AA (1978). Studies on the genus *Euphorbia* L. in Egypt. II. Systematic treatment. Taeckholmia 9:9-57.
- El Hadidi MN, Fayed AA (1995). Materials for excursion flora of Egypt (EFE). Taeckholmia 15:1-233.
- Fossati J, Pautou G, Peltier JP (1998). Wadi vegetation of the North-Eastern desert of Egypt. Feddes Repertorium 109:313-327.
- Galal TM, Fahmy AG (2011). Plant diversity and community structure of Wadi Gimal protected area, Red Sea Coast of Egypt. African Journal of Ecology 50:266-276.
- Gitay H, Noble IR (1997). What are functional types and how should we seek them?, p 3-19. In: Smith TM, Shugart HH, Woodward FI (Eds.). Plant functional types. Cambridge University Press, Cambridge.
- Gomaa NH (2012). Soil seed bank in different habitats of the Eastern Desert of Egypt. Saudi Journal of Biological Sciences 19:211-220.
- Gouvas M, Theodoropoulos K (2007). Life-form and chorological spectra of the vegetation units of Mount Hymettus (C Greece). Journal of Biological Researchthessaloniki 8:177-187.
- Hassib M (1951). Distribution of plant communities in Egypt. Bulletin de l'Institut du Desert d'Egypte 29:60-261.
- Hegazy A, Al-Atar AA, Lovett-Doust J, El-Adawy H (2012). Spatial and temporal plant phenological niche differentiation in the Wadi Degla desert ecosystem (Egypt). Acta botanica Croatica 71(2):261-277.
- Henderson PA, Seaby RMH (1999). Community Analysis Package (CAP) version 1.2. Pisces Conservation Ltd. IRC House, UK.
- Hillel D, Tadmor J (1962). Water regime and vegetation in the central Negev highlands of Israel. Ecology 43:33-41.
- Jackson ML (1967). Soil Chemical Analysis-Advanced Course, Department of Soil Sciences, Washington.
- Jongman RH, Ter Braak CJF, van Tongeren OFG (1987). Data analysis in community and landscape ecology. Pudoc, Wageningen.
- Kapur P, Govil SR (2000). Experimental Plant Ecology 1st Ed. CBS Publishers and Distributors, New Delhi.
- Kassas M (1966). Plant life in deserts, p. 145-180. In: Hills ES (Eds.). Arid Lands, London: Mathuen, Parism, UNESCO.
- Kassas M, Girgis WA (1965). Habitat and plant communities in the Egyptian desert. VI. The units of desert ecosystem. Journal of Ecology 53:719-729.
- Kassas M, Zahran MA (1962). Studies on the ecology of the Red Sea coastal land. I. The district of Gebel Ataqa and El-Galala El-Bahariya. Bulletin Society Geographie d'Egypte 35:129-175.
- Klimes L (2003). Life-forms and clonality of vascular plants along an altitudinal gradient in E Ladakh (NW Himalayas). Basic and Applied Ecology 4:317-328.
- Lyon J, Sagers CL (2003). Correspondence analysis of functional groups in a riparian landscape. Plant Ecology

164:171-183.

- Mahdavi P, Akhani H, van der Maarel E (2013). Species diversity and life form patterns along 3000 m altitudinal gradient in the steppe vegetation of southern slopes of the Alborz Mountains, Iran. Folia Geobotanica 48:7-22.
- Monod TH (1954). Mode contracte 'et diffus de la vegetation Saharienne. p. 35-44. In: Cloudsley-Thompson JL (Eds.). Biology of Desert, Institute of Biology, London.
- Moustafa AA, Klopatek JM (1995). Vegetation and landforms of the Saint Catherine area, southern Sinai, Egypt. Journal of Arid Environment 30:385-395.
- Moustafa AA, Zaghloul MS (1996). Environment and vegetation in the montane Saint Catherine, south Sinai, Egypt. Journal of Arid Environment 34:331-349.
- Noy-Meir I (1973). Desert ecosystems: higher trophic levels. Annual Review of Ecology, Evolution, and Systematics 5: 195–214
- Olsvig-Whittaker L, Shachak M, Yair A (1983). Vegetation patterns related to environmental factors in a Negev Desert watershed. Vegetatio 54:153-165.
- Orlóci L (1978). Multivariate Analysis in Vegetation Research, 2nd Ed. Dr. W. Junk, The Hague.
- Quézel P (1978). Analysis of the flora of Mediterranean and Saharan Africa. Annals of the Missouri Botanical Garden 65:479-534.
- Ryan PJ, McKenzie NJ, Loughhead A, Ashton L (1996). New methods for forest soil surveys. In: The role of *Eucalyptus* and other fast growing species. (Eds.). Eldridge KG, Crowe MP, Olds KM. Csiro Publishing, Collingwood, Victoria.
- Said R (1962). The Geology of Egypt, Elsevier, Amsterdam, 377 p.
- Salama FM, Abd El-Ghani MM, El-Naggar SM, Baayo K (2005). Vegetation structure and environmental gradients in the Sallum area, west Mediterranean, Egypt. Ecologia Mediterranea 31(1):15-32.
- Salama FM, Abd El-Ghani MM, El-Tayeh N (2013). Vegetation and soil relationships in the inland wadi ecosystem of central Eastern Desert, Egypt. Turkish Journal of Botany 37:489-498.
- Salama FM, Ahmed MK, El-Tayeh NA, Hammad SA (2012). Vegetation analysis, phenological patterns and chorological affinities in Wadi Qena, Eastern Desert, Egypt. African Journal of Ecology 50:193-204.
- Salama FM, El-Naggar SM (1991). Phytosociology of wadi system west of Qusseir province. Feddes Repertorium 102:453-468.
- Salama FM, Fayed AA (1990). Phytosociological study on the deltaic part and the principal channel of wadi Qena, Egypt. Feddes Repertorium 101:89-95.
- Shaheen AM (2002). Weed diversity of newly farmed land on the southern border of Egypt (Eastern and Western shores of Lake Nasser). Pakistan Journal of Biological Sciences 5(7): 802-806.
- Shaltout KH, El-Halawany EF, El-Garawany MM (1997). Coastal lowland vegetation of eastern Saudi Arabia.

57

Biodiversity and Conservation 6:1027-1070.

- Sheded MG, Radwan UA, Taher MA, Springuel I (2012). Spatial heterogeneity in hyper-arid vegetation of the South Western Desert, Egypt. Feddes Repertorium 122(5-6):351-366.
- Shmida A (1985). Biogeography of the desert flora, p. 149– 202. In: Evenari M, Noy-Meir I, Goodall DW (Eds.). Ecosystems of the world 12 B: Hot deserts and arid shrublands, Elsevier, Amsterdam.
- Smith T, Huston M (1989). A theory of the spatial and temporal dynamics of plant communities. Vegetatio 83:49-69.
- Springuel I, Ali MM (1990). Impact of Lake Nasser on desert vegetation in desert development. Proc 2 International Desert Development Conference, Cairo, 557-568.
- Springuel I, El-Hadidi MN, Sheded M (1991). Plant communities in southern part of the Eastern Desert (Arabian Desert) of Egypt. Journal of Arid Environment 21:307-317.
- Springuel I, Sheded MG, Darius F, Bornkamm R (2006). Vegetation dynamics in an extreme desert wadi under the influence of episodic rainfall. Polish Botanical Studies 22:495-472.

Stahr K, Bornkamm R, Gauer A, Kehl H (1985). Veränderung

von Boden und Vegetation am Übergang von Halbwüste zur Vollwüste zwischen Mittelmeer und Quattare Depression in Ägypten. Geoökodynamik 6:99-120.

- Suzan AS, Gadallah MAA, Salama FM (2013). Ecophysiological studies on three desert plants growing in Wadi Natash, Eastern Desert, Egypt. Journal of Biology and Earth Sciences 3(1):135-143.
- Täckholm V (1974). Student's Flora of Egypt, 2nd Ed. Cairo University Press, Cairo, 888 p.
- Tielbörger K (1997). The vegetation of linear desert dunes in the north-western Negev, Israel. Flora 192:261-278.
- White F (1993). Long distance dispersal and the origins of the afromontane flora. Sonderb. Naturwiss. Vereins Hamburg 7:87-116.
- White F, Léonard J (1991). Phytogeographical links between Africa and Southwest Asia. Flora et Vegetatio Mundi 9:229-246.
- Yair A, Danin A (1980). Spatial variation as related to the soil moisture regime over an arid limestone hillside, northern Negev, Israel. Oecologia 47: 83-88.
- Zahran MA, Willis AJ (2009). The Vegetation of Egypt, 2nd Ed. Springer Science and Business Media B.V., London, 541 p.