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Intracellular Biosynthesis and Antibacterial Activity of Silver Nanoparticles Using Edible Mushrooms

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Abstract

The process of biosynthesis of silver nanoparticles is a simple, cost effective and eco-friendly approach. Biosynthesis of silver nanoparticles using some commonly available edible mushroom extracts and their antimicrobial activity was demonstrated in the current study. The formation of silver nanoparticles was confirmed by UV, FTIR and SEM and antibacterial activity was tested using disc diffusion method. From the results it is confirmed the successful formation of silver nanoparticles using mushroom extracts; they performed their role as a reducing and capping agent and also exhibited a potent antibacterial activity against *S. aureus* (gram positive bacteria). Thus the biosynthesis of silver nanoparticles using edible mushroom extract will deserve to be a good candidate as an antibacterial agent.

Keywords: antibacterial activity, biosynthesis, edible mushrooms, silver nanoparticles

Introduction

Despite the fact of advances in developing new drugs against various infectious diseases, till date there is a growing concern about the emergence and re-emergence of drug resistance. Currently nanotechnology has paved way in developing novel drugs mainly in the field of medicine. The use of nanoparticles is presently gaining impetus as they posses defined chemical, optical and mechanical properties. They are most promising as they show good antibacterial properties due to their large surface area to volume ratio, which is coming up as the current interest in the researchers, due to the growing microbial resistance against metal ions, antibiotics and the development of resistant strains (Gong et al., 2007). Different types of nanomaterials like copper, zinc, titanium (Retchkiman-Schares et al., 2006), magnesium, gold (Gu et al., 2003), alginate (Ahmad et al., 2005) and silver have recently come up for the synthesis of silver nanoparticles. Among other noble metals, silver nanoparticles have proved to be most effective as it has good antimicrobial efficacy against bacteria, viruses and other eukaryotic micro-organisms. Since ancient times silver has been used for burn wounds treatment, dental work, catheters and bacterial infection control, in forms of metallic silver, silver nitrate, and silver sulfadiazine Previously, nanoparticles were synthesized only by physical and chemical methods. Nowadays researchers are showing much interest on biological mediated synthesis using plants, fungi, microbes and yeast (Philip, 2009; Tripathy

et al., 2009). Extracts from bio-organisms may act both as reducing and capping agents in Ag NPs synthesis. The reduction of Ag+ ions by combinations of biomolecules found in these extracts, such as enzymes/proteins, amino acids, polysaccharides, and vitamins, is environmentally benign, yet chemically complex. This was achieved either through intracellular and extracellular synthesis (Acosta, 2009).

Mushrooms have been part of the normal human diet for thousands of years and in recent times, the amounts consumed have risen greatly, involving a large number of species. Generally edible mushrooms are used for nutritional and therapeutic purposes (Borchers et al., 2004; Chang, 1996; Ribeiro and Salvadori, 2003). Apart from this, they are valuable health foods since they are low in calories, fats, essential fatty acids and high in vegetable proteins, vitamins and minerals (Murugkar and Subbulakshmi, 2005). Among various mushrooms Calocybe indica (milky mushroom) [CI], Pleurotus ostreatus (oyster mushroom) [PO], Agaricus bisporous (button mushroom) [AB] and Ganoderma lucidum (reishi mushroom) [GL] which are commonly available edible mushrooms, revealed to possess wide beneficial properties such as antioxidant, anticancer, antimicrobial (Lillian et al., 2008). Due to its increased nutritive and therapeutic values herein we focussed on the simple and ecofriendly approach to synthesis silver nanoparticles using the above mentioned edible mushrooms and further evaluated its antibacterial activity.

Materials and methods

Chemicals

All chemicals were of analytical grade and procured from Sigma Aldrich (India) or Merck (India) unless otherwise stated. The culture media were purchased from Hi-Media (India).

Preparation of intracellular mushroom extract

20 g of fruiting bodies of edible mushrooms (PO, AB, GL and CI) were taken separately and rinsed thrice in distilled water, dried on a tissue paper and cut in fine pieces until they were made into a paste, and finally boiled in 100ml of sterile distilled water up to 5 minutes. It was filtered using whatman No: 41 filter paper and stored at 4°C.

Intracellular Biosynthesis of Silver Nanoparticles

1 mM aqueous solution of silver nitrate (AgNO3) was prepared and used for the synthesis of silver nanoparticles. 10 ml of mushroom extract was added into 90 ml of aqueous solution of 1 mM silver nitrate for reduction into Ag+ ions and incubated overnight at room temperature in dark.

UV- Vis Spectral Analysis of Silver Nanoparticles

The reduction of pure Ag+ ions was monitored by measuring the UV-vis spectrum of the reaction medium after overnight incubation, after diluting a small aliquot of the sample into distilled water. Silver nanoparticles (Ag-NPs) are soluble in distilled water and the colour changes were observed visually. A yellowish brown colouration was noticed at the synthesis phase. The concentration of AgNP produced was measured using a Systronics UV double-beam spectrophotometer (model 2201), at a resolution of 1 nm, between 200 and 600 nm, using 10 -mm -optical- path- length quartz cuvettes.

Fourier Transform Infrared Spectroscopy (FTIR)

FT-IR spectra were measured using Nicolet-Avatar 300 FT-IR spectrometer. About 5 mg of sample were mixed with 100 mg of KBr and compressed into pellet using a hydraulic press. FT-IR spectra over the scanning range of 4000-400 cm⁻¹ were obtained with the resolution of 2 cm⁻¹.

Preparation of Silver Nanoparticles Sample for SEM Studies

The morphological features of synthesized nanoparticles were examined by scanning electron microscopy (JEOL 6380A, Japan).

Antibacterial activity of Silver nanoparticles

Antibacterial activity of medicinal compounds is treated through Disc diffusion methods, most commonly employed method to evaluate the antibacterial activity. In the present study we have used the disc diffusion method to check the antibacterial activity of mushroom extracts. The disc diffusion techniques are widely used in Kirby-Bauer method.

Disc diffusion was carried out for the bacterial suspension containing 10^6 cells. Plates with Nutrient agar were seeded and then drained off. They were desiccated at room temperatute for 15-20 min for adequate drying. The disc prepared from fruiting bodies extract was placed in quadrangular manner in different petri dishes. Then petri dishes were incubated at $30\pm2^{\circ}$ C for 24 hours. After 24-48 hours results were noted for the zone of inhibition, which was measured in mm.

Results and discussion

The current study focused on some edible mushrooms which mediated the synthesis of silver nanoparticles and also investigated for its antimicrobial activity. Even though several approaches have been reported to synthesize Ag-NPs, currently biological mediated synthesis has received great interest in the scientific community. Since mushrooms are highly enriched with proteins it is suspected that they act as reducing, as well as capping, agent.

Biosynthesis of silver nanoparticles

Synthesis of silver nanoparticles using PO, AB, GL and CI extract was observed (Fig. 1). When mushroom extract was individually subjected to aqueous solution of silver nitrate, a gradual change of colour was observed after 48 hrs. PO, AB and GL extract changed its colour to reddish brown, while CI extract changed its colour to yellow, indicating the formation of silver nanoparticles after 48 hrs. This change of colour could be due to the formation of silver nanoparticles with the recent report of Philip (2009). The formation of reduced AgNPs reaction mixture was further characterised by UV-vis spectrophotometry.

Generally, silver nanoparticles absorb in the visible region of the electromagnetic spectrum at 380-450 nm, due to the surface plasmon resonance (SPR) transition. In the present study the reduction of silver ions to silver nanoparticles using PO, AB and GL mushroom extracts was completed within 48 hrs whereas the reduction of the reaction mixture for CI extract was completed within 24 hrs and there was no further reduction. The nanoparticles synthesized using PO extract were absorbed at 420 nm, with AB extract at 410 nm, with GL extract it was absorbed at 430 nm and CI extract was absorbed at 400 nm. Thus all the nanoparticles synthsized using mushroom extract showed the absorption spectra between 400-430 nm, which indicates the presence of silver nanoparticles, which is due to the excitation of surface plasmon resonance in the metal nanoparticles (Mulvaney, 1996). After 48 hrs of incubation, no further increase of the intensity was recorded, indicating complete reduction of precursor silver ions. This formation of AgNPs was mediated by the active biomol-

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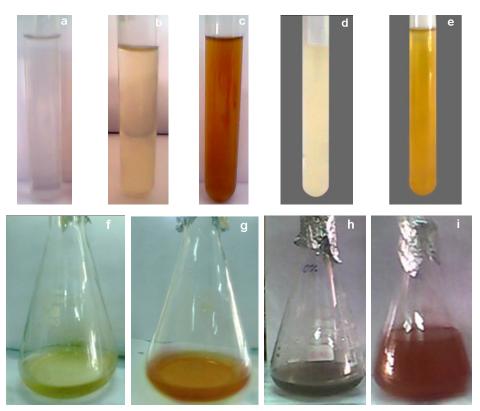


Fig. 1. Optical photograph of a) AgNO₃, b) PO, c) PO+AgNo₃, d) AB, e) AB+AgNo₃, f) GL, g) GL+AgNo₃, h) CI, i) CI+AgNo₃ before and after 48 hrs of reaction

ecules present in the mushroom extracts. Strictly pursuing Philips *et al.* (2011), has also reported the biosynthesis of AgNPs using edible mushroom which strongly supports our result.

FTIR analysis of PO and PO-AgNPs

FTIR measurements showed the evidence of possible interactions between silver and bioactive molecules of mushroom extract, which may be responsible for the synthesis and stabilization (capping material) of silver nanoparticles. Fig. 2 shows the FTIR spectra of *i*) PO extract and PO-AgNPs, *ii*) AB extract and AB-AgNPs, *iii*) GL extract and GL-AgNPs, *iv*) CI and CI-AgNPs. The FTIR peaks determined the bonds relevant to amine N-H stretching, aldehyde C-H stretching, bending vibration of amide I and II, carboxylic acid stretching and aromatic ring stretching. Silver nanoparticles usually absorb strongly at 1679, 1539, 1452, 1430 and 1040 cm⁻¹ (Justin Packia Jacob *et al.*, 2012).

In this study PO-AgNPs showed intense absorption at 1649, 1544, 1459, 1397, 1082 cm⁻¹ which is assigned to be amide I and II group and C-N stretching vibrations of aromatic ring; this strongly proves the association of PO in reducing silver ions. In the spectra of AB and AB-AgNPs exhibited vibration at 1635 cm⁻¹ and 1545 cm⁻¹ found in AB-AgNPs are the bending vibrations of the amide I and II, which clearly indicates the presence of proteins in the *A. bisporous* mushroom extract. The bands observed at 1320,

1243 and 1047 cm⁻¹ can be assigned to the C-N stretching vibrations of aromatic and aliphatic amines respectively. GL extract showed the absorption peak at 3433 cm⁻¹, which is assigned to O-H stretching of alcoholic groups. The peak at 1742 cm⁻¹ can be assigned an absorption peaks of C=O, while the peak at 1629 cm⁻¹ corresponds to N-H bending vibrations of the amide I and II, which clearly indicates the presence of proteins in the G. lucidum extract. The absorbance peak at 1358, 1326, 1210 and 1054 cm⁻¹ corresponds to C-N stretching respectively. CI extract exhibited vibration at 3418 cm⁻¹ and 3422 cm⁻¹, which is assigned to the N-H group from peptide linkage present in the extract. 1641 cm⁻¹ and 1550cm⁻¹ absorbed in CI-AgNPs are the bending vibrations of the amide I and II, which clearly indicates the presence of proteins in the C. indica mushroom extract. CI-AgNPs showed intense absorption at 1641, 1550, 1458, 1387, 10782 cm⁻¹, which strongly proves the association of C. indica in reducing silver ions.

These variations of N-H, O-H and other infrared bands, suggests the formation of silver nanoparticles using PO, AB, GL and CI extract. It is well known that these edible mushrooms posses various phytochemicals like proteins, polysaccharides and amino acids which have acted as a bioreductant for the biosynthesis of silver nanoparticles. It is also possible that proteins/enzymes play a role in the reduction of metal ions by the oxidation of benzaldehyde

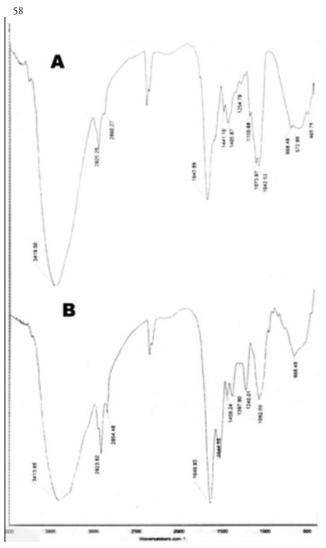


Fig. 2. *i*). FTIR spectrum of A) PO and B) PO-AgNPs

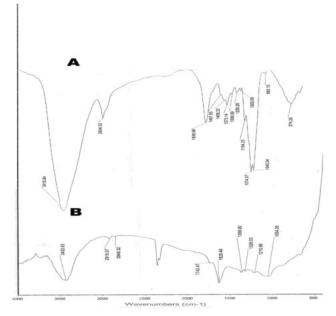


Fig. 2. *iii*). FTIR spectrum of A) GL and B) GL-AgNPs

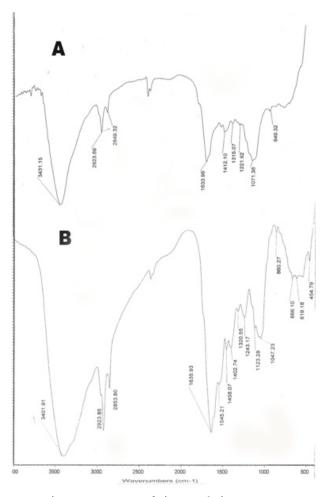


Fig. 2. *ii*). FTIR spectrum of A) AB and B) AB-AgNPs

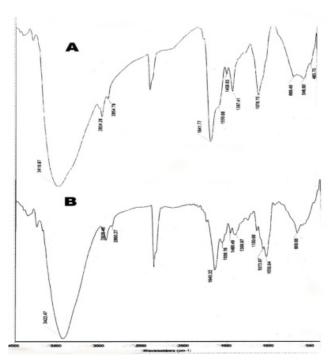


Fig. 2. *iv*). FTIR spectrum of A) CI and B) CI-AgNPs

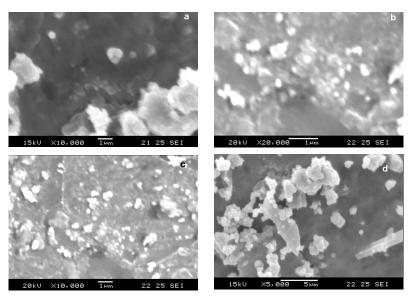


Fig. 3. SEM images of a) PO-AgNPs, b) AB-AgNPs, c) GL-AgNPs, d) CI-AgNPs

(aldehyde groups) to carboxylic acids (Mandal *et al.*, 2006; Philip, 2009; Shankar *et al.*, 2004).

SEM analysis

The morphology and the particle size of Ag NPs synthesized using PO, AB, GL and CI extract was determined using SEM micrograph (Fig. 3). NPs synthesized using PO seems to be spherical with agglomerated particles and the sizes found are roughly in the range of 100 nm. SEM micrograph of AB-Ag NPs showed to be spherical in aggregated form and its size range was of 80-100 nm. GL-Ag NPs was spherical in shape, with its size around 50 nm. CI-Ag NPs was also found to be spherical in shape, with its size range around 100 nm, thus proving it to be a nanoparticle.

Antibacterial activity

Bacterial clinical isolates normally exhibit higher levels of resistance to antibiotics, as they are under continuous selective antibiotic stress in medical care facilities (Rahman *et al.*, 2012). Moreover, bacterial strains associated with food-related outbreaks may also demonstrate higher levels of resistance, likely due to the use of antibiotics in animal husbandry (Smith *et al.*, 2002). It is widely proved that silver nanoparticles have showed good antibacterial activity against different bacterial strains (Bankura *et al.*, 2012). Numerous reports have stated the antibacterial activity of Ag NPs (Hindi *et al.*, 2009; Lansdown, 2002; Morones *et al.*, 2005). In the present study the antibacterial activity of silver nanoparticles was investigated against pathogenic bacteria of gram positive strains using disc diffusion method. The antibacterial activity of synth-

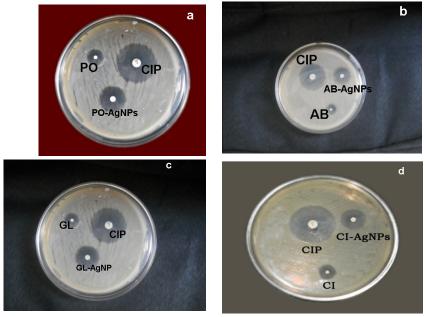


Fig. 4. Antimicrobial activity of silver nanoparticles against S. aureus (gram positive)

sized PO-AgNPs, AB-AgNPs, GL-AgNPs and CI-AgNPs against gram positive (Staphylococcus aureus) bacteria was evaluated using standard zone of inhibition (Fig. 4 and Tab. 1). Ciproflaxacin was used as a reference antimicrobial drug. The diameter of inhibitory zones on both free extract and AgNPs were represented in Tab. 1. PO and GL AgNPs showed maximum zone of inhibition at 7 mm and 8 mm, where as for AB and CI AgNPs the inhibitory zone was around 6 mm. In the case of mushroom extract the inhibitory zone was less when compared to their respective AgNPs, which proved to have less antimicrobial activity. From our results we suggest that AgNPs exhibited antibacterial activity against gram positive bacteria while the exact mechanism of the antibacterial effect of silver ions is not exactly known. The antibacterial activity is probably derived through the electrostatic attraction between negative charged cell membrane of microorganism and positive charged nanoparticles (Dibrov et al., 2002). Shrivastava et al. (2007), studied antibacterial activity against E. coli and S. aureus. This potential antimicrobial activity of silver nanoparticles has made them promising candidates as an antimicrobial agent.

Conclusion

Synthesis of silver nanoparticles using various edible mushrooms extracts was developed in a very simple and eco-friendly method. Generally, mushrooms containing proteins have played a major role in acting as a reductant as well as a capping material in order to synthesize a novel AgNPS and functioned more effectively as an antimicrobial agent against gram positive bacterial strains. Thus, the prospect of rational use of mushrooms in synthesizing metallic nanoparticles is an exciting and very simple biological method which will prompt with high outlook in future, especially in various medicinal fields.

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