

Effects of Salinity and $\text{NO}_3:\text{NH}_4$ Ratio on Yield and Quality in Canola (*Brassica napus* L.)

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Abstract

The effect of salinity and $\text{NO}_3:\text{NH}_4$ ratio (0:100, 75: 25, 50:50 and 25:75) in the nutrient solution on growth, yield quality and N metabolism in hydroponically grow canola (*Brassica napus* L.) was evaluation. Both fresh and dry weights of leaves were significantly lower when a high concentration of either NO_3 (100%) or NH_4 (75%) was the sole N source in the nutrient solution. In nonsaline condition, increasing of both NH_4 and NO_3 ratio in the nutrient solution reduced photosynthetic (pn) rate, however in salinity condition the reduction of pn became more pronounced at a higher ratio of NH_4 in the nutrient solution. The yield in terms of fresh and dry weight of seed per plant was significantly increased at the 75:25 ($\text{NO}_3:\text{NH}_4$) treatments. Total fat in nonsaline condition was increased with increasing NH_4 ratio in the nutrient solution, however in saline condition it was reduced, at high NH_4 ratio in the nutrient solution. The increase of tissue N concentration was nearly proportional to the NH_4 concentration in the nutrient solution. The activity of nitrate reductase (NR) was increased by increasing NH_4 form 0 to 50% and then reduced at a higher ratio of NH_4 in the solution. Salinity increased NH_4 concentration so that the saline condition had nearly twice high NH_4 concentration in the leaves. The increase of NH_4 concentration induced by salinity could be partially the reduction of NH_4 assimilate because of the shortage of carbohydrate.

Keywords: nitrogen form, nutrient solutions, salinity, photosynthesis, canola

Introduction

Plants take up nitrogen mainly as nitrate and ammonium. When both are present in solution ammonium uptake is preferred. Under salt stress, nitrate uptake is slowed down and salinity reduces NO_3 assimilation with the possible consequence of N deficient in the plant (Oliel *et al.*, 2005). The $\text{NO}_3:\text{NH}_4$ ratio in the root zone offers an important means for the controlling of the relative uptake on nutrients (Sonneveld, 2002). Although the major form of N supply in the nutrient solution in the soil-less culture is NO_3 based fertilizer, addition of some NH_4 to the nutrient solution seems to be beneficial to the plant (Gashaw and Mugwira, 1981) (Sonneveld *et al.*, 1999), and helps to decrease the pH of the solution (Ikeda and Osawa, 1983). The studies show that the response of plants to either NO_3 or NH_4 concentration varies with the species, temperature, and salinity (Edwards and Horton, 1982). The assimilation of NO_3 usually occurs mostly in the leaves of plants, however the stage of NO_3 assimilation seem to be linked closely enough to preclude accumulation of NH_4 (Pilbeam and Kirkby, 1992). Modification of $\text{NH}_4:\text{NO}_3$ ratio in the nutrient solution modulates the relative uptake of anions and cations (Clay *et al.*, 1984). The increase of NH_4 to NO_3 ratio in the root zone impairs growth and

reduces the yield (Guo *et al.*, 2002). The presence of high concentration of NH_4 has been shown to induce a decreased concentration of cations such as K, Ca and Mg, while NO_3 has the opposite effect (Alan, 1989; Kotsiras *et al.*, 2002). Under intensive fertigation, nitrogen not only affects plant growth, it may also alter the salinity tolerance of plants depending on its ionic form. The beneficial effects of nitrate under saline conditions have been attributed to the antagonism between NO_3 - and Cl ions (Feigin, 1990). The balance of the cation to anion uptake ratio by secretion of H^+ , HCO_3^- , or organic anion from root cells also has an effect on the rhizosphere. Adjusting the $\text{NO}_3:\text{NH}_4$ ratio from the total cation to anion uptake ratio and maintain pH within desired range (Kafkafi *et al.*, 1982).

Whereas growth suppression by ammonium probably results from a shortage of sugar in the roots (Cramer *et al.*, 1995) or from inhibition of nitrate reductase activity in roots and shoots (Botell *et al.*, 1993). Some reports, however, indicate that combined NH_4 and NO_3 fertilization at an 50:50 ratio results in greater biomass accumulation compared with plants fed with either nitrogen source alone (Lips *et al.*, 1990). In canola, some of the damaging effects of salinity have been averted by partial substitution if nitrate with ammonium in the nutrient solution. In tomatoes, improved plant growth and higher yield were

obtained with addition of low levels of NH_4 to the mainly nitrate containing fertigation solution (Flores *et al.*, 2001). Therefore, the objective of this experiment was to assess the effect of combination of salinity and NO_3 : NH_4 ratio on the growth yield and N metabolism in canola.

Materials and methods

The experiment was conducted at agricultural research center of Tabriz, Iran. Seedling of canola (*Brassica napus* L. var SLM046,) were planted in the propagation pots. When two leaves were fully expanded, the seedling were transferred to the glasshouse and placed into pots filled with a mixture of perlite and vermiculite (1:1, v/v). For each saline and nonsaline treatment, the experimental design was a completely randomized design with four replicates per treatment. The greenhouse was under natural sunlight during spring and summer and the temperature was set to $22^\circ\text{C}\pm 3$ and $15^\circ\text{C}\pm 3$ day and night, respectively. The plants were subjected to two salinity levels (0, 200 mM NaCl). The plants in different saline conditions were fed with one of the following NO_3 : NH_4 ratio : 0:100, 75:25, 50:50 and 25:75. Modifying Hoagland's solution (Hoagland and Arnon, 1950) was used to prepare the nutrient solution. The concentration of the nutrients in the solutions was as follow (in mg.l^{-1}): 300K, 170 Ca, 50 Mg, 60 S, 33 P, 1.5 B, 0.1 Cu, 2 Mn, 3Zn, 12 Fe (Fe-DTPA), and 0.1 Mo. Nitrogen at 200 mg.l^{-1} was provided NO_3 and NH_4 forms to give NO_3 : NH_4 ratio of 100:0, 75:25, 50:50 and 25:75 (Tab. 1). To obtain identical K, Ca, Mg, total N and P concentrations in all treatments, the changes in NO_3 : NH_4 ratios were balanced by varying the Cl concentration. The electrical conductivity (EC) of the nutrient solution was within the range 2.7-2.8 dS.m^{-1} . The balance of NO_3 : NH_4 ratio changes by varying the concentration of at least one more nutrient was inevitable due to the charge balance constrain (Schrevens and Cornell, 1993; Savvas *et al.*, 2003). However difference in EC within these ranges are unlikely to influence significantly plant growth (Sonneveld *et al.*, 1999). The initial pH of

the nutrients solutions containing NO_3 and NH_4 was adjusted to 6.5- 6.8 by adding H_2SO_4 (Adams, 2002).

The Ec and pH of drainage water from pots were checked every week in the drainage water from the pots and an additional solution was applied to minimize Ec and pH changes in the root zone. The concentration of Ca and K in the plant tissues was measured by atomic absorption spectrophotometry (Perkin Elmer, Model 110, USA). The concentration of N and P in the youngest fully expanded leaves were determined by kjeldahl and colorimetric method, respectively. The activating of nitrate reductase (NR) in the young leaves samples was measured according to Gebauer *et al.*, (1998) with some modification. At maturity all plants were harvested and seed yield per plant was recorded for each treatment. For the extraction of oil from seed, the dried seeds were ground to a uniform powder and oil was extracted in n-hexane. A weighed quantity (15g) of the seed powder was placed in an extraction thimble and extraction was carried out for 6 hr in a Soxhlet extractor, after which time the oil containing solvent was filtered through whatman No.42 filter paper. The oil was heated in an oven at 60°C to remove last traces of the solvent. The leaf area was measured using a leaf area- meter (Licor, Model Li-1300, USA). After being weighed, the leaves were dried at 80°C in an air-forced oven for 48.h. The photosynthetic (Pn) rate of the mid- lamina portion of the youngest fully expanded leaves (two leaves) from each experimental unit was measured with a portable photosynthesis-meter (Walz, Model Da-1010, Germany) from 9:00 to 12 :00 a.m. Reference CO_2 concentration was set to the inside glasshouse. The rates of Pn were measured at $450 \mu\text{l.l}^{-1}$ CO_2 concentration and $800 \mu\text{mol.m}^{-2}\text{S}^{-1}$ PAR. The leaf chambers temperature was set up to 28°C and relative humidity was ranged from 70 to 80%. The data were subjected to analysis of variance (ANOVA) using generalized linear models by using the SAS 8.2 software package. The significance of the treatment effects presented as , not significant; * $P<0.05$, ** $P<0.01$. In case of significant treatment effects, comparison of means was performed using LSD at a significance level of 0.05.

Tab. 1. The concentrations of salts (mM) used to prepare nutrient solutions at NO_3 : NH_4 ratio of 100:0, 75:25, 50:50 and 25:75.

| Salts | NO_3 : NH_4 ratios in the solutions | | | |
|------------------------------------|---|---------|---------|---------|
| | 100 : 0 | 75 : 25 | 50 : 50 | 25 : 75 |
| KNO_3 | 5.7 | 5.4 | 0.0 | 0.0 |
| $\text{Ca}(\text{NO}_3)_2$ | 4.3 | 2.7 | 3.6 | 1.8 |
| MgSO_4 | 2.0 | 2.0 | 2.0 | 2.0 |
| KH_2PO_4 | 1.0 | 0.0 | 0.0 | 0.0 |
| $\text{NH}_4\text{H}_2\text{PO}_4$ | 0.0 | 1.0 | 1.0 | 1.0 |
| NH_4Cl | 0.0 | 2.6 | 6.2 | 9.8 |
| KCl | 1.0 | 2.3 | 7.7 | 7.7 |
| CaCl_2 | 0.0 | 1.5 | 0.7 | 2.4 |

Results and discussion

The vegetative characteristics as a function of various N forms in the nutrient solution at the salinity levels of canola plants are given in Tab. 2. Both fresh and dry weight of leaves were significantly lower when a high concentration of either NO_3 (100%) or NH_4 (75%) was the sole N Source in the solution. The highest both leaf number and leaf area were observed in 25:75 and 50:50 (NO_3 : NH_4) treatment. In both saline and nonsaline condition, the index of chlorophyll was increased with increasing NH_4 ratio in the solution.

Within each column, same letters indicate no significant difference between treatment ($P<0.05$).

Tab. 2. The effects of salinity and $\text{NO}_3:\text{NH}_4$ ratio on the vegetative characteristics of canola plants

| Salinity | $\text{NO}_3:\text{NH}_4$ ratio | Leaf fresh weight (g.plant ⁻¹) | Leaf dry weight (g.plant ⁻¹) | Leaf area (cm ²) |
|-------------------------------|---------------------------------|--|--|------------------------------|
| Non-saline | 100:0 | 273.8 b | 39.3 b | 1876 b |
| | 75:25 | 286.5 a | 43.6 a | 1988 a |
| | 50:50 | 288.7 a | 44.8 a | 1996 a |
| | 25:75 | 274.4 b | 38.7 b | 1867 b |
| Saline condition (200mM NaCl) | 100:0 | 169.3 c | 36.9 c | 8372 c |
| | 75:25 | 183.2 b | 39.3 b | 9928 b |
| | 50:50 | 185.1 b | 43.7 | 1160 ab |
| | 25:75 | 170.3 c | 35.5 d | 7687 d |

There was a significant reduction of vegetative characteristic of canola plant in saline condition (Tab. 2). Leaf area in saline condition was only 84% of that in nonsaline condition. In nonsaline condition, the increased both NH_4 and NO_3 ratio in the nutrient solution reduced pn rate. In saline conditions, the reduction in pn rate was severely impaired at a higher ratio of NH_4 in the nutrient solution. The yields in terms of fresh weight per plant were significantly increased at the 75:25 and 50:50 ($\text{NO}_3:\text{NH}_4$) treatment (Tab. 2). Both high ratios of NH_4 and NO_3 reduced fresh weight per plant. Fresh and dry weight of seeds in the saline condition was reduced by 25% and 29.5%, respectively, as compared to nonsaline condition. Fat percent was increased in nonsaline condition in (50:50), (75:25) ($\text{NO}_3:\text{NH}_4$) treatments Higher availability of carbohydrate was the result of the increased pn rate and fat. The concentration of Ca in the seeds was significantly reduced in 25:75 ($\text{NO}_3:\text{NH}_4$) treatment (Tab. 3). The data of leaf ions concentration of the plants relative to salinity and ($\text{NO}_3:\text{NH}_4$) ratio are presented in (Tab. 3).

Within each column, same letters indicate no significant difference between treatment ($P < 0.05$).

The increase of tissue N concentration in the nutrient solution. Both high concentrations of NO_3 was observed in 75:25 and 100:0 ($\text{NO}_3:\text{NH}_4$) treatments. Salinity significantly increased NH_4 concentration. High ratio of NH_4 in the nutrient solution and salinity increased Ca concentration in the leaf tissue (Tab. 4). The activity of

NR was increased by increasing NH_4 from 0 to 50% and then reduced at a higher ratio of NH_4 in the solution (Tab. 4). The highest NR activity was observed in 50:50 ($\text{NO}_3:\text{NH}_4$) treatment. The vegetative characteristics of canola were affected by the ratio of $\text{NO}_3:\text{NH}_4$ in the nutrient solution and salinity. Increasing concentration of both NH_4 and NO_3 significantly reduced the canola plant nonsaline condition. This reduction became more severe in saline condition. There was a reduction in the leaf area (40.5%) in nonsaline and saline plants when grown in 75% NH_4 compared with plants grown in 50% NH_4 our results are disagreement with the earlier report by Claussen (2002) and Tabatabaei *et al.* (2006) who showed that the NH_4 form of N caused a decrease in dry weight in tomato, but led to an increase in dry weight when supplied besides NO_3 .

Within each column, same letters indicate no significant difference between treatment ($P < 0.05$).

This experiment clearly demonstrated that both high concentration of NO_3 and NH_4 has an inhibitory effect on growth of canola and it becomes more pronounced in the salinity conditions. Many researchers (Gashaw and Mugwira, 1981; Salsac *et al.*, 1987) have reported the beneficial effects of two forms of N in an appropriate ratio on the growth of different plants. Less growth of plants fed with high concentration of NH_4 may be due to the unavailability of NO_3 as a N source and higher demand of carbohydrates channeled for NH_4 assimilation and detoxification. On the other hand, complete exclusion of NH_4 from the solution reduced the canola plant growth. Both fresh weight and reduced under 100:0 ($\text{NO}_3:\text{NH}_4$) and 25:75 ($\text{NO}_3:\text{NH}_4$) ratio and became more pronounced in salinity. The possible explanation is that high concentration of NH_4 reduces the uptake of cations such as Ca and K and increase the concentration of NH_4 , which is toxic for the plant (Sasseville and Mills, 1979; Alan, 1989). The higher reduction of fresh weight in saline conditions may be due to increased NH_4 concentration and reduced carbohydrate level.

It is well known that salinity reduces photosynthesis, carbohydrate levels, and plant weight in various of species of crop.

Tab. 3. The effects of salinity and $\text{NO}_3:\text{NH}_4$ ratio on the concentration of mineral nutrients

| Salinity | $\text{NO}_3:\text{NH}_4$ ratio | NO_3 (mg.g ⁻²) | N(mg.g ⁻¹) | P(mg.g ⁻¹) | K(mg.g ⁻¹) | NH_4 (mg.g ⁻¹) |
|-------------------------------|---------------------------------|-------------------------------------|------------------------|------------------------|------------------------|-------------------------------------|
| Non-saline | 100:0 | 3.7 a | 32.8 c | 3.0 bc | 44.0 b | 1.7 c |
| | 75:25 | 3.6 a | 34.6 b | 6.0 a | 47.0 a | 1.9 b |
| | 50:50 | 2.5 b | 36.4 a | 9.0 a | 48.5 a | 2.5 b |
| | 25:75 | 2.3 b | 39.4 a | 8.0 a | 41.3 b | 3.6 a |
| Saline condition (200mM NaCl) | 100:0 | 4.5 a | 32.4 c | 2.5 c | 64.5 a | 2.3 b |
| | 75:25 | 3.8 a | 34.6 b | 4.5 b | 57.5 a | 2.9 b |
| | 50:50 | 3.8 a | 40.4 a | 4.0 b | 50.2 a | 4.2 a |
| | 25:75 | 2.8 b | 42.4 a | 9.8 a | 44.6 b | 4.5 a |

Tab. 4. The effects of salinity and NO₃: NH₄ ratio on pn, Ca concentration and Nitrate reductas activity

| Salinity | NO ₃ : NH ₄ ratio | Photosynthetic Rate (μMol.m ⁻² s ⁻¹) | Ca concentration (mg.g ⁻¹) | Nitrate reductase activity | Activity fat (%) |
|-------------------------------|---|---|--|----------------------------|------------------|
| Non-saline | 100:0 | 10.4 b | 4.3 d | 1.8 c | 38 bc |
| | 75:25 | 12 ab | 9.2 b | 2.4 bc | 40 ab |
| | 50:50 | 13.3 a | 8.5 bc | 3.2 ab | 41 a |
| | 25:75 | 10 b | 3.8 d | 1.6 c | 39 b |
| Saline condition (200mM NaCl) | 100:0 | 6.4 d | 7.2 c | 2.7 b | 35 |
| | 75:25 | 8.2 c | 11.7 a | 2.8 b | 38 bc |
| | 50:50 | 9.5 bc | 10.7 ab | 3.8 a | 39 b |
| | 25:75 | 6.2 d | 6.4 c | 1.7 c | 34 c |

Nova and loomis (1981) proposed that the demand for N is determined by growth rate and the N composition of new tissue. Because of the relation between N form and photosynthesis, it has been suggested that the N concentration of leaf is adjusted to the irradiance experienced during growth to make full use of the intercepted radiation. In this experiment, the decrease of dry weight in both saline and nonsaline conditions was nearly proportional to the leaf area, as total photosynthesis sources, in higher NH₄ concentration in the nutrient solution. Leaves are the main photoassimilates sources for the plant (Taiz and Zeiger, 2002) and the biomass production is associated with both leaf area and the pn rate. A reduction in pn rate at the high ratio of both NH₄ and NO₃ concentration occurred.

This result agrees with the findings of Takacs and Tecs (1992), who reported a reduction of pn in NH₄ fed plants. The increased activity of pn enzymes with NH₄ as opposed to NO₃ nutrition is generally the results of increased protein content of NH₄ fed plants (Golvano *et al.*, 1982). Some researchers have argued that the accumulation of NH₄ in the leaves may cause uncoupling of the electron transport from photosynthesis in chloroplast, resulting in decreasing photosynthetic rate (Puritch and Baker, 1967; Claussen 1986; Rothstein and Cregg, 2005).

The inhibition of pn by toxicity of NH₄ has been reported by many researchers (Givan, 1979; Frantz *et al.*, 1982). The higher demand for carbon skeletons to detoxify excess NH₃ in plants supplied with NH₄ as sole N source imposes an opposing effect on plant growth (Beevers and Hag eman, 1969). Moreover, at high external NH₄ concentration, the uptake of N predominantly in cationic form suppresses the total anion uptake; this process is balanced by excessive H⁺ secretion from the root cells (Kirkby and Knight, 1977; Pilbeam and Kirkby, 1992). Consequently, the pH in the root environment may decline to level that are detrimental for most plant species. Although the reduction in pn at high NH₄ concentration has been reported in many plants, the reason for the reduced pn at high concentration of NO₃ has not been found. It seems that higher concentration of NO₃

has an inhibitory effect on pn rate. It is well known that highest growth rates are archived when an adequate supply balanced the sucrose supply of amino acids. This means that under good pn conditions a high N supply along with proper N form is required for best growth. Fresh and dry weight were in nonsaline conditions in 50:50(NO₃:NH₄) treatment. Higher availability of carbohydrate was the result of the increased pn rate and higher leaf area led to better growth of nonsaline condition in 50:50 (NO₃:NH₄) treatment. Lower yield by plants under saline rendered low photosynthetic activity. This became more severest the increased concentration of NH₄ in the root zone (Claussen and lenz, 1999, Grindly, 1997).

Both salinity and the ratio of NO₃:NH₄ in the nutrient solution altered the nutrient concentration in the leaves. The greater concentration of total N and NH₄ was observed with increasing NH₄ concentration in the nutrient solution. This is consistent with the finding Lorenzo *et al.* (2000). Who reported that the addition of ammonium in a nutrient solution produced as total nitrogen uptake increase during shoot elongation of rose, while without ammonium, nitrate uptake was lower during shoot elongation. Unfavorably high concentration of NH₄ in the nutrient solution may lead to NH₄ toxicity, which considered to be the result of effects such as NH₄ induced nutrient deficiency caused by impaired uptake of ions, acidification of the root zone, alteration in the osmotic balance, modification of phytohormone, and impaired of N enzyme metabolism (Gerendas *et al.*, 1997; Lorenzo *et al.*, 2000). It is well known that NH₄ nutrition leads to the acidification of the root zone, however in this experiment the pH of solution was controlled to eliminate this factor as an impact on plant growth. Plants take up nitrogen mainly as nitrate and ammonium. (Mengel and Viro 1978; Sawas, *et al.*, 2003). NH₄ also reduces the uptake of Ca and K by the roots. The activity of NR in the canola plants was affected by NO₃: NH₄ ratio. An important prerequisite of adaptation of plant species and cultivars to certain environmental conditions are enzymes that are capable for catalyzing nitrogen reduction and assimilation (Lee and Stewart; 1978, Runge, 1983). The activity of NR has often been shown to

decrease dramatically if plants were fed with ammonium instead of nitrate nitrogen (Claussen, 1986). A positive correlation between NR activity and NO_3^- concentration in the leaf tissues implies that the activity of NR is induced by increasing NO_3^- concentration, as it has been reported by Sivaasankar and Oaks (1996).

Conclusions

The data present in this paper suggest that salinity has effected on the yield of canola and its effects is modulated by N form in the nutrient solution. Use of 50:50 or 75:25 ($\text{NO}_3^-:\text{NH}_4^+$) ratio in root to cause increasing yield of canola plant in saline and nonsaline conditions.

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