

Emelike CU *et al.* (2023) **Notulae Scientia Biologicae** Volume 15, Issue 3, Article number 11315 DOI:10.15835/nsb15311315 Research Article



# Modulation of antioxidant activities, markers of hepatic and renal dysfunctions in alloxan-induced diabetic rats by *Combretum dolichopetalum*

# Chinedum U. EMELIKE<sup>1\*</sup>, Ojichukwuka E. CHIJIOKE-AGU<sup>1</sup> Favour-Ann K. NWOKE<sup>1</sup>, Romanus OYIBE<sup>1</sup>, Chinwendu G. ENWEREUZO<sup>2</sup>, Eze F. AHUEKWE<sup>3</sup>

 <sup>1</sup>Alex Ekwueme Federal University, Department of Physiology, Faculty of Basic Medical Sciences, Ndufu-Alike, Nigeria; chinedum.emelike@funai.edu.ng (\*corresponding author); melaninojii@gmail.com; favour.nwoke@funai.edu.ng; oyiberomanusagbo@gmail.com
<sup>2</sup>Middlesex University, Department of Cardiology, Faculty of Science and Technology, The Burroughs, NW4 4BT, Hendon, London, United Kingdom; ce476@live.mdx.ac.uk
<sup>3</sup>Covenant University, Department of Biological Sciences, College of Science and Technology, Ota, Nigeria; eze.ahuekwe@covenantuniversity.edu.ng

# Abstract

Diabetes mellitus is a metabolic dysfunction of insulin secretion exhibiting hyperglycemia with abnormalities in protein, fat and carbohydrate metabolism. The present study aimed to investigate the effect of *Combretum dolichopetalum* (CD) on hepatic parameters, renal indices and antioxidant markers of alloxaninduced diabetic rats. Twenty (20) male rats were used, and they were divided into four groups of five rats each. Group 1- Normal control received distilled water (non-diabetic control); Group 2- Diabetic control received distilled water; Group 3 and Group 4 are Diabetic rats treated with 200 mg/kg and 400 mg/kg of CD extract. There were significant decreases in the final body weights of Group 2 compared with Group 1 and an elevation of the body weights of Groups 3 and 4 compared with Group 1 and a reduction in blood glucose concentration of Group 2 compared with Group 2. There were significant increases in the alanine amino transaminase (ALT), alkaline phosphatase (ALP) activities and serum total and conjugated bilirubin levels of Groups 2 and 3 compared with Group 1, but a reduction in the ALP, ALT activities and total bilirubin in Group 4. There were reductions in catalase activities and no change in the renal indices. The study showed ameliorating potentials on hyperglycemia, hepatoprotective activity and antioxidation effects.

Keywords: Combretum dolichopetalum; diabetes mellitus; metabolic disorder; oxidative stress

*Received: 19 Jul 2022. Received in revised form: 13 Jul 2023. Accepted: 25 Aug 2023. Published online: 04 Sep 2023.* From Volume 13, Issue 1, 2021, Notulae Scientia Biologicae journal uses article numbers in place of the traditional method of continuous pagination through the volume. The journal will continue to appear quarterly, as before, with four annual numbers.

# Introduction

Diabetes mellitus is a looming health challenge that has taken the global health sector by storm. It is a functional flaw of the human body and loses its ability to convert sugar (glucose) into energy for consumption and use by the body (Ogbera and Ekpebegh, 2014). This condition is a metabolic dysfunction depicting chronic hyperglycemia with irregularities in the metabolism of protein, fat and carbohydrates arising from flaws around insulin secretion, action or both (ADA, 2009). Diabetes mellitus has proven to be responsible for long term damage, dysfunction and failure of vital organs. Hepatic and renal dysfunction appears almost simultaneously concerning individual independent organ failures or a collective of organ failures in fatally ill patients (Darmon *et al.*, 2014). These dysfunctions emerge simultaneously in disease cases involving the liver and kidney, primary hepatic dysfunction with secondary renal dysfunction and primary renal dysfunctions is ready to share similar pathogenetic mechanisms.

The liver is the body's detoxification organ and plays an essential role in maintaining metabolic homeostasis. Illnesses associated with the liver are of great essence because of this specific function (Kalra *et al.,* 2023). Free radicals produced from compounds are metabolised in the liver. These free radicals are further scavenged by antioxidants to maintain oxidative/antioxidative balance in the liver. However, when the oxidative/antioxidative balance scale is overturned, which leads to a condition termed "Oxidative Stress". Oxidative stress leads to damaging processes in the liver and ends up in liver diseases (Muriel and Arauz, 2012).

The use of medicinal plants in the treatment and prevention of specific illnesses/ailments have demonstrated a vital role in the global healthcare system (Dasgupta and Bratati, 2007; Okoro *et al.*, 2021; Ekakitie *et al.*, 2021; Aloke *et al.*, 2021a; Aloke *et al.*, 2021b; Aloke *et al.*, 2021c; Emelike *et al.*, 2021). Phenolics have the capacity to scavenge free radicals resulting from their redox properties, commonly found in leaves, flowering tissues and woody parts such as stems and barks of plants (Larson, 1988).

*Combretum dolichopetalum* is an indigenous African plant predominantly found in the Eastern parts of Nigeria. The plant serves a vital role in food nutrition and is also significant in ethnomedicine. In food nutrition, the leaves are boiled and used as soup (Ameyaw *et al.*, 2012; Uzor *et al.*, 2014; Barku *et al.*, 2014; Emelike *et al.*, 2020). The study of the medicinal properties of plants to bring relief to ailments has evolved. It delivers the arsenal required to fight off oxidative stress-related diseases such as hepatic and renal dysfunctions, cancer, cardiovascular diseases and other neurodegenerative disorders. It has become predominant due to the assemblage of free radicals in the body (Sahoo *et al.*, 2012).

The present study aimed to determine the modulation of antioxidant activities, markers of hepatic and renal dysfunctions in alloxan-induced diabetic rats by *Combretum dolichopetalum*.

#### Materials and Methods

#### Plant materials

Fresh matured leaves of *C. dolichopetalum* were located and collected in 2019 from its natural habitat in Nsukka, Enugu. Mr C. J. Onyeukwu, a taxonomist of the Department of Plant Science and Biotechnology, University of Nigeria, authenticated the plant. A voucher specimen (UNH No.49a) was deposited at the herbarium.

# Preparation of extract

The leaves extract was washed and air-dried at room temperature for 7 days. It was grounded to a coarse powder using an electric blender (model ms-233, China). About two kilograms (2 kg) were extracted for 48 h with methanol in a Soxhlet extraction following the method of Jensen (2007). Following the extraction, the

extract was collected and dried at a low temperature (40 °C) to obtain the pale dark green used for animal experiments.

# Chemicals

Alloxan monohydrate used in this study was sourced from Sigma-Aldrich Chemical Company, United States of America. All other chemicals employed were of standard grade.

# Animal experiments

Twenty (20) male rats weighing about 120-200 g were used for the study obtained from the Animal House, Department of Physiology, University of Nigeria Enugu Campus. They were acclimatized to their feed (Vital feed<sup>®</sup>, Nigeria) and water (which they had access to *ad libitum*) for two weeks before the commencement of the experiment. The study protocol was approved by the College of Medicine Research Ethics Committee of the University, with protocol number 026/02/2017. The study followed the established institutional guidelines and the NIH guidelines for experimental animals.

# Induction of experimental diabetes mellitus

After two (2) weeks of acclimatization, a freshly prepared solution of alloxan monohydrate (0.5 g dissolved in 8.5 ml of distilled water) was administered intraperitoneally to fifteen (15) rats at a dosage of 150 mg/kg body weight at a fasting state. The remaining five (5) rats served as the non-diabetic control group. Blood samples were collected from the tail vein of the rats for blood glucose concentration analysis using a blood glucometer (Accu-Chek \*, India) before the commencement of the administration of the extract. The alloxan-treated rats with fasting blood glucose levels>200 mg/dl after seven days of induction and evidence of hyperglycemia considered to be diabetic were used for the study.

### Experimental procedure

The normal rats and rats with stable diabetes mellitus were assigned into four groups of five rats per group.

The groups were as follows:

Group 1- Normal control received distilled water (non-diabetic control)

Group 2- Diabetic control also received distilled water.

Group 3- Diabetic rats treated with 200 mg/kg of CD extract.

Group 4- Diabetic rats treated with 400 mg/kg of CD extract.

Bodyweight (before and after administration) was measured using a digital electronic weigh Scout Pro SP 401 (China). At the end of twenty-eight (28) days of the administration, the rats fasted overnight, and blood was collected from their tail for blood glucose analysis using a blood glucose meter. The rats were anaesthetized with 2% sodium pentobarbital (75 mg/kg) intraperitoneally. Venous blood was obtained via the orbital, poured into plain tubes and allowed to clot. Sera were obtained from the clotted sample after centrifuging at 3000 rpm for 10 minutes for the analysis of antioxidant activity and liver and renal function parameters.

#### Body weights

The changes in the weights of the rats were recorded using a digital electronic weighing scale model number Scout Pro SP 401 (China).

## Liver function tests

The liver indices analysed include total bilirubin and direct bilirubin. Others include alanine aminotransferase (ALT), aspartate aminotransferase (AST) and alkaline phosphatase (ALP) using A25 Biosystem (Barcelona, Spain).

# Kidney function tests

Electrolytes (sodium, potassium, chloride and bicarbonate) were estimated with Easylyte<sup>®</sup> analyzer Medica Corporation, Bedford, USA while urea and creatinine were analysed with A25 Biosystem, Barcelona, Spain.

# Antioxidant activity

The level of superoxide dismutase (SOD) was measured following the procedure of Assady *et al.* (2011). The activity of catalase (CAT) in serum was determined according to the method described by Hadwan (2018). The serum concentration of reduced glutathione (GSH) was measured by the method described by Alisik (2019).

# Statistical analysis

Results were analysed using GraphPad Prism (GraphPad<sup>\*</sup> Software, San Diego, CA, USA). One-way ANOVA following the Turkey post hoc test was used for data comparison. The values were regarded as significant at P<0.05. Results were expressed as mean  $\pm$  standard error of the mean

# Results

#### Effect of C. dolichopetalum on body weight of rats

Table 1 shows the body weight and percentage change in the body weights of the rats. There were no significant differences (P>0.05) in the initial body weights of all the rats in the respective groups and no significant differences (P>0.05) in the final body weights of Groups 3 and 4 compared with Group 1.

In contrast, there were significant decreases (P<0.05) in the final body weights of Group 2  $(24.7\pm1.00$  loss of weight) compared with the Group 1 but significant elevation (P<0.05) of the final body weights of Groups 3 and 4 compared with the Groups 2.

Parameters	Group 1	Group 2	Group 3	Group 4
Initial body weight (g)	$178.2 \pm 2.00$	179.1±1.90	180.2±2.50	180.3±1.00
Final body weight (g)	205.1±2.30	157.0±1.30*	197.3±1.40#	206.2±3.40#
Weight difference (g)	26.3±2.10	$24.7 \pm 1.00$	$14.7 \pm 1.00$	24.0±2.89
	(increase)	(decrease)	(increase)	(increase)

Table 1. Effect of C. dolichopetalum on body weight of rats

Values are mean  $\pm$  SEM, n=5, \*= P< 0.05 versus normal control group #= P< 0.05 versus diabetic control

## Effect of C. dolichopetalum on blood glucose concentration of rats

Table 2 shows the blood glucose concentration of the rats that were studied. There were significant differences (P<0.05) in the blood glucose concentration of Groups 2, 3 and 4 compared with Group 1 in the initial blood glucose. Also, there were significant increases (P<0.05) in the final blood glucose concentration of Group 2 compared with Group1, but a significant reduction (P<0.05) of their hyperglycemia in Groups 3 and 4 compared with Group 2.

Parameters	Group 1	Group 2	Group 3	Group 4
Initial glucose level (mg/dL)	$76.2 \pm 5.03$	299.1±1.40*	$308.2 \pm 4.50^*$	$314.0 \pm 4.80^*$
Final glucose level (mg/dL)	$100.1 \pm 4.30$	306.0±3.30*	174.3±1.50*#	113.2±3.10*#
Glucose difference (mg/dL)	$24.3 \pm 2.10$	26.7±3.30	133.4±5.90	201.0±6.30
	(increase)	(increase)	(decrease)	(decrease)

Table 2. Effect of C. dolichopetalum on blood glucose concentration of rats

Values are mean  $\pm$  SEM, n=5, \*= P< 0.05 versus normal control group #= P< 0.05 versus diabetic control

# Effect of C. dolichopetalum on the liver function test of rats

The serum AST activities of Group 3 were significantly (P<0.05) increased when compared with Group 1. There were significant increases (P<0.05) in the ALP, ALT activities and Serum Total and Conjugated bilirubin levels of Groups 2 and 3 compared with Group 1. Also, there were significant increases (P<0.05) in the ALP and ALT activities of Group 3 when compared with Group 2 but a significant reduction (P<0.05) of the ALP, ALT activities and Total bilirubin in Group 4 when compared with Group 2 (Table 3).

Table 3. Effect of *C. dolichopetalum* on the liver function test of rats

Parameters	Group 1	Group 2	Group 3	Group 4
AST (IU/L)	32.90±1.69	40.90±2.33	49.47±2.78*	26.63±1.98
ALP (IU/L)	110±3.40	127±2.40*	204±6.50*#	96.9±2.40#
ALT (IU/L)	26.2±2.39	38.9±3.32*	92.3±4.22*#	21.7±1.07#
Total bilirubin (μmol/L)	9.50±0.93	19.3±0.77*	23.7±2.90*	10.6±2.35#
Conjugated bilirubin (µmol/L)	2.20±0.15	3.10±0.12*	3.37±0.15*	2.50±0.17

Values are mean  $\pm$  SEM, n=5, \*= P < 0.05 versus normal control group #= P < 0.05 versus diabetic control

# Effect of C. dolichopetalum on renal indices of rats

There was no significant (P>0.05) change in the renal indices of Groups 2, 3 and 4 when compared with Group 1 (Table 4).

Parameters	Group 1	Group 2	Group 3	Group 4
Sodium (mmol/L)	$140 \pm 2.03$	143±0.58	$142 \pm 1.00$	$144 \pm 0.88$
Potassium (mmol/L)	12.7±3.61	10.6±1.82	$11.2 \pm 1.44$	$7.27 \pm 1.84$
Chloride (mmol/L)	$101 \pm 2.08$	103±0.58	102±1.15	$104 \pm 0.58$
Bicarbonate (mmol/L)	24.3±1.20	25.0±1.15	22.3±0.88	24.7±0.33
Urea (mmol/L)	6.43±0.79	9.63±2.10	$9.0 \pm 1.74$	$8.47 \pm 1.98$
Creatinine (mmol/L)	94.0±8.66	89.3±7.8	86.3±3.9	86.0±3.00

Table 4. Effect of C. dolichopetalum on renal indices of rats

Values are mean  $\pm$  SEM, n=5, \*= P < 0.05 versus normal control group <sup>#</sup>= P < 0.05 versus diabetic control

# Effect of C. dolichopetalum on antioxidant of rats

The oxidative stress markers of the rats are shown in Table 5. There were no significant differences (P>0.05) in the SOD activities of Groups 2 and 3 compared with Group 1. On the other hand, there was a significant elevation (P<0.05) in the SOD activity of Group 4 compared with Groups 1 and 2. There were significant reductions (P<0.05) in the catalase activities of Groups 2 and 3 compared with Group 1.

Parameters	Group 1	Group 2	Group 3	Group 4
SOD (U/M)	9.46±0.41	8.92±0.97	11.2±0.55	12.6±0.39*#
GPx (U/M)	$0.80 \pm 0.14$	$1.28 \pm 0.14$	$0.97 \pm 0.11$	$0.96 \pm 0.12$
CAT (KU/L)	65.9±2.6	46.7±2.31*	39.7±0.26*	57.9±6.20

Table 5. Effect of C. dolichopetalum on antioxidant of rats

Values are mean  $\pm$  SEM, n=5, \*= P< 0.05 versus normal control group \*= P< 0.05 versus diabetic control

#### Discussion

In the present study, there was an increase in the body weight of Group 1 (Normal control), which suggests the increased synthesis of tissue proteins (Eleazu *et al.*, 2019). The reduction in the body weight in the diabetic control group (Group 2) could be a result of alloxan destroying the pancreatic cells leading to insulin deficiency, which causes raised production of ketone bodies. Thus, elevated ketone bodies with increased lipolysis result in a loss in body weight (Cotter *et al.*, 2013). However, the gain in body weight observed in diabetic rats treated with *C. dolichopetalum* could be attributed to better modulation of hyperglycemia in the diabetic rats and reduction in fasting blood glucose which could improve body weight in alloxan-induced diabetic rats (Yin *et al.*, 2018). Additionally, the ability of the *C. dolichopetalum* extracts to enhance body weight may be due to its ability to lower elevated blood glucose via increased glucose metabolism, and this may be attributed to the protective effect of the extract at a higher dose. This is in agreement with the report of Eleazu *et al.* (2014).

The alloxan monohydrate rat model of diabetes is one of the most commonly used because it mimics many of the complications of human diabetes (Szkudelski, 2001). There is increasing evidence that alloxan causes diabetes by rapid depletion of  $\beta$ -cells by DNA alkylation and accumulation of cytotoxic free radicals resulting from initial islets inflammation and infiltration of activated macrophages and lymphocytes in inflammatory focus. This will reduce plasma insulin concentration, resulting in a sustained hyperglycemia state (Szkudelski, 2001). The reduction of the blood glucose concentrations of diabetic rats following the administration of *C. dolichopetalum* to the extent observed in this study indicates the ameliorating potentials of *C. dolichopetalum* on hyperglycemia. The hypoglycemic effect of the extracts may be attributed to the enhanced secretion of insulin from the beta cells of the pancreas or may be due to increased tissue uptake of glucose by enhancement of insulin sensitivity (Maniyar and Bhixavatimath, 2012; Emelike *et al.*, 2020). Many researchers have reported that flavonoids are potent bioactive antioxidants and anti-diabetic agents and that the alkaloid content of plants could regulate insulin secretion. Also, saponins have been shown to exhibit blood-glucose-lowering potentials (Patel *et al.*, 2012; Emelike *et al.*, 2021).

The liver is necrotic in diabetic rats, which will lead to increased activities of total bilirubin, conjugated bilirubin, ALP, ALT and AST (Aloke *et al.*, 2021b). They leak from the liver cytosol into the bloodstream. It is also an indicator of the hepatotoxicity of alloxan (Saeed *et al.*, 2008). It is in agreement with this present study. However, the oral administration of *C. dolichopetalum* at 400 mg/kg suggests a non-deleterious effect on the total bilirubin and serum liver enzymes. The liver is an essential organ that helps in drug metabolism and other toxicants. The destruction of the liver cell results in the impairment of the liver cell membrane permeability, which results in the leakage of tissue contents into the bloodstream (Saeed *et al.*, 2008). Hyperglycemia increases the generation of free radicals by glucose auto-oxidation, and the increment of free radicals may lead to liver damage (Tangyarasittichai, 2015).

The non-significant effect *C. dolichopetalum* had on the kidney indices of the normal rats, diabetic rats and diabetic treated rats suggests the non-deleterious effect of *C. dolichopetalum* on the kidney.

Oxidative stress is involved in the pathogenesis of many forms of genetic and acquired hypertension (Griendling *et al.*, 2013). Poorly managed diabetes frequently results in nephropathy and cardiovascular complications (Bramlage *et al.*, 2019). Moreover, the alleviation of oxidative stress with antioxidant therapy has been shown to ameliorate hypertension in several animal models (Griendling *et al.*, 2013). Studies have shown that uncontrolled hyperglycemia in rats was associated with the activity of antioxidant enzymes (Eleazu *et al.*, 2014). However, administration of *C. dolichopetalum* at doses of 200 and 400 mg/kg increased SOD compared to Group 1 (Normal control). 400 mg/kg of *C. dolichopetalum* significantly increased SOD compared to Diabetic control (Group 2). It suggests that an increase in SOD activity is probably due to the regeneration of the damaged liver cells by the *C. dolichopetalum* extract and the viability of diabetic rats to secret

insulin. The effects of the *Combretum dolichopetalum* extract were dose-dependent. Furthermore, this potent antioxidant activity could be attributed to the phenolic content of the *Combretum dolichopetalum*.

#### Conclusions

The study showed ameliorating potentials on hyperglycemia, hepatoprotective activity and antioxidation effects.

# Authors' Contributions

Conceptualization: CUE, RO and CGE; Data curation: CUE, RO and CGE; Formal analysis: CUE, RO and CGE; Investigation: CUE, RO and CGE; Methodology: CUE, OEC and FKN; Project administration: EFA; Resources: CUE, RO and CGE; Software: EFA; Supervision: CUE; Validation: CUE; Visualization: CUE, RO, CGE, OEC and FKN; Writing - original draft: CUE and OEC; Writing - review and editing: CUE, FKN and EFA. All authors read and approved the final manuscript.

# Ethical approval (for researches involving animals or humans)

The study protocol was approved by the College of Medicine Research Ethics Committee of the University, with protocol number 026/02/2017.

# Acknowledgements

This research received no specific grant from any funding agency in the public, commercial, or not-forprofit sectors.

# **Conflict of Interests**

The authors declare that there are no conflicts of interest related to this article.

# References

- Alisik M, Neselioglu S, Erel O (2019). A colorimetric method to measure oxidized, reduced and total glutathione levels in erythrocytes. Journal of Laboratory Medicine 43(5):269-277. *https://doi.org/10.1515/labmed-2019-0098*
- Aloke C, Emelike CU, Obasi NA, Ogbu PN, Edeogu CO, Uzomba CG, ... Bature CU (2021a). HPLC profiling and studies on *Copaifera salikounda* methanol leaf extract on phenylhydrazine-induced hematotoxicity and oxidative stress in rats. Arabian Journal of Chemistry 14(12):103428. https://doi.org/10.1016/j.arabjc.2021.103428
- Aloke C, Nwachukwu N, Obasi NA, Emelike CU, Amu PA, Ogbu PN, Orinya OF, Ogbonnia EC (2021b). Antihyperglycemic, antihyperlipidemic and hepatoprotective effects of *Ficus ottoniifolia* (Miq.) Miq. supplementation in alloxan-induced diabetic rats. Avicenna Journal of Phytomedicine 11(5):428-435. https://doi.org/10.22038/AJP.2020.16958

- Aloke C, Obasi NA, Emelike CU, Ogbu PN, Ufebe GO, Orinya OF, Egwu CO, Onyekwere AC (2021c). Protective effect of *Copaifera salikounda* (Heckel) against paracetamol-induced hepatorenal injury in rat. Sains Malaysiana 50(4):1065-1076. http://doi.org/10.17576/jsm-2021-5004-17
- American Diabetes Association (2009). Diagnosis and classification of diabetes mellitus. Diabetes Care 32(1):S62-S67. https://doi.org/10.2337/dc09-S062
- Ameyaw Y, Barku VYA, Ayivor J, Forson A (2012). Phytochemical screening of some indigenous medicinal plant species used in the management of diabetes mellitus in Ghana. Journal of Medicinal Plants Research 6:4573-4581. https://doi.org/10.5897/JMPR12.564
- Assady M, Farahnak A, Golestani A, Esharghian M (2011). Superoxide dismutase (SOD) enzyme activity assay in *Fasciola* spp. parasites and liver tissue extract. Iranian Journal of Parasitology 6(4):17-22. https://pubmed.ncbi.nlm.nih.gov/22347309/
- Barku VYA, Boahen O, Dali G (2014). Ethnobotanical study of wound healing plants in Kpando traditional area, Ghana. International Journal of Phytomedicine (6):564-572. http://aj.yloop.com/index.php/ijpm/article/view/1519/0
- Bramlage P, Lanzinger S, van Mark G. Hess E, Fahrner S, Heyer CHJ, ... Holl RW (2019). Patient and disease characteristics of type-2 diabetes patients with or without chronic kidney disease: an analysis of the German DPV and DIVE databases. Cardiovascular Diabetology 18:33. https://doi.org/10.1186/s12933-019-0837-x
- Cotter DG, Schugar RC, Crawford PA (2013). Ketone body metabolism and cardiovascular disease. American Journal of Physiology-Heart and Circulatory Physiology 304(8):H1060-1076. https://doi.org/10.1152/ajpheart.00646.2012
- Darmon M, Clec'h C, Adrie C, Argaud L, Allaouchiche B, Azoulay E, Bouadma L, ... Timsit JF (2014). Acute respiratory distress syndrome and risk of AKI among critically ill patients. Clinical Journal of the American Society of Nephrology 9(8):1347-1353. https://doi.org/10.2215/CJN.08300813
- Dasgupta N, Bratai De (2007). Antioxidant activity of some leafy vegetables of India: A comparative study. Food Chemistry 101(2):471-474. https://doi.org/10.1016/j.foodchem.2006.02.003
- Ekakitie O, Okoro FE, Nwite EJ, Chukwunweike C, Fredrick CC, Emelike CU (2021). Methanolic extract of *Citrullus lanatus* seeds abates testicular degeneration and dose-dependently modulates testicular function in hyperlipidemic male Wistar rats. Nigerian Journal of Physiological Sciences: Official Publication of the Physiological Sciency of Nigeria 36(1):101-107.
- Eleazu C, Ekeleme CE, Famurewa A, Mohamed M, Akunna G, David E, ... Emelike U (2019). Modulation of the lipid profile, hepatic and renal antioxidant activities, and markers of hepatic and renal dysfunctions in alloxan-induced diabetic rats by virgin coconut oil. Endocrine, Metabolic and Immune Disorders Drug Targets 19(7):1032-1040. https://doi.org/10.2174/1871530319666190119101058
- Eleazu CO, Eleazu KC, Chukwuma SC, Okoronkwo J, Emelike CU (2014). Effect of Livingstone potato (*Plectranthus esculenthus* N. E. Br) on hyperglycemia, antioxidant activity and lipid metabolism of streptozotocin induced diabetic rats. Toxicology Reports 1:674-681. https://doi.org/10.1016/j.toxrep.2014.08.013
- Emelike CU, Anyaehie USB, Iyare EE, Obike CA, Aloke C, Chukwu DF, ... Chukwu JAO (2021). Chemical composition and evaluation of methanol leaf extract of *Combretum dolichopetalum* on body weights and haematological indices of phenylhydrazine induced-anaemic rats. Toxicology International 28(2):8-14.
- Emelike CU, Anyaehie USB, Iyare EE, Obike CA, Eleazu CO, Chukwuma C (2020). Acute and sub-acute toxicity studies on *Combretum dolichopetalum* Engl. and Diels leaves. Slovenian Veterinary Research 57(3):105-114. https://doi.org/10.26873/SVR-899-2020
- Griendling KK, Camargo LL, Rios FJ, Alves-Lopes R, Montezano AC, Touyz RM (2021). Oxidative stress and hypertension. Circulation Research 128(7):993-1020. *https://doi.org/10.1161/CIRCRESAHA.121.318063*
- Hadwan MH (2018). Simple spectrophotometric assay for measuring catalase activity in biological tissues. BMC Biochemistry 19(1):7. https://doi.org/10.1186/s12858-018-0097-5
- Jensen WB (2007). The Origin of Soxhlex Extractor. Journal Chemistry Education 84(12):1913-1914. https://doi.org/10.1021/ed084p1913
- Kalra A, Yetiskul E, Wehrle CJ, Tuma F (2023). Physiology. Liver. In: StatPearls. Treasure Island (FL): StatPearls Publishing *https://pubmed.ncbi.nlm.nih.gov/30571059/*
- Larson RA (1988). The antioxidants of higher plants. Phytochemistry 27:969-978. https://doi.org/10.1016/0031-9422(88)80254-1

- Maniyar Y, Bhixavatimath P (2012). Antihyperglycemic and hypolipidemic activities of aqueous extract of *Carica papaya* Linn. leaves in alloxan-induced diabetic rats. Journal of Ayurveda and Integrative Medicine 3(2):70-74. https://doi.org/10.4103/0975-9476.96519
- Moore K, Taylor G, Ward P, Williams R (1991). Aetiology and management of renal failure in acute liver failure. In Williams R, RD Hughes. Acute liver failure: Improved understanding and better therapy. Welwyn Garden City, UK: Smith Klein and French Laboratories pp 47-53
- Muriel P, Arauz J (2012). Coffee and liver health. In: Chu Y (Ed). Coffee emerging health effects and disease prevention. West Sussex, UK: IFT Press/Wiley-Blackwell pp 123-139
- Ogbera AO, Ekpebegh C (2014). Diabetes mellitus in Nigeria: The past, present and future. World Journal of Diabetes 5(6):905-911. *https://doi.org/10.4239/wjd.v5.i6.905*
- Okoro CO, Aloke C, Ibiam UA, Obasi NA, Orji OU, Egwu CO, Ogbu PN, Emelike CU, Ufebe GO, Ezeani NN (2021). Studies on ethanol extracts of *Olax subscorpioidea* against carbon tetrachloride-induced hepatotoxicity in rats. Pakistan Journal of Biological Science 24(6):724-732.
- Patel DK, Prasad SK, Kumar R, Hemalatha S (2012). An overview on antidiabetic medicinal plants having insulin mimetic property. Asian Pacific Journal of Tropical Biomedicine 2(4):320-230. https://doi.org/10.1016/S2221-1691(12)60032-X
- Saeed MK, Deng Y, Dai R. (2008). Attenuation of biochemical parameters in streptozotocin-induced diabetic rats by oral administration of extracts and fractions of *Cephalotaxus sinensis*. Journal of Clinical Biochemistry and Nutrition 42:21-28.
- Sahoo S, Ghosh G, Das D, Nayak S (2012). Phytochemical investigation and in vitro antioxidant activity of an indigenous medicinal plant *Alpinia nigra* B.L.Brutt. Asian Pacific Journal of Tropical Biomedicine 3(11):871-876. https://doi.org/10.1016/S2221-1691(13)60171-9
- Szkudelski T (2001). The mechanism of alloxan and streptozotocin action in B cells of the rat pancreas. Physiological Research 50(6):537-546.
- Tangvarasittichai S (2015). Oxidative stress, insulin resistance, dyslipidemia and type 2 diabetes mellitus. World Journal of Diabetes 6(3):456-480. https://doi.org/10.4239/wjd.v6.i3.456
- Uzor PF, Osadebe PO, Omeje EO, Agbo MO (2014). Bioassay guided isolation and evaluation of the antidiabetic principles of *Combretum dolichopetalum* root. Journal of Pharmaceutical Research International 18(4):2155-2171. https://doi.org/10.9734/BJPR/2014/9143
- Yin P, Wang Y, Yang L, Sui J, Liu Y (2018). Hypoglycemic effects in alloxan-induced diabetic rats of the phenolic extract from Mongolian oak cups enriched in ellagic acid, kaempferol and their derivatives. Molecules 23(5):1046. https://doi.org/10.3390/molecules23051046



The journal offers free, immediate, and unrestricted access to peer-reviewed research and scholarly work. Users are allowed to read, download, copy, distribute, print, search, or link to the full texts of the articles, or use them for any other lawful purpose, without asking prior permission from the publisher or the author.

**License** - Articles published in *Notulae Scientia Biologicae* are Open-Access, distributed under the terms and conditions of the Creative Commons Attribution (CC BY 4.0) License.

© Articles by the authors; Licensee SMTCT, Cluj-Napoca, Romania. The journal allows the author(s) to hold the copyright/to retain publishing rights without restriction.

#### Notes:

- Material disclaimer: The authors are fully responsible for their work and they hold sole responsibility for the articles published in the journal.
- Maps and affiliations: The publisher stay neutral with regard to jurisdictional claims in published maps and institutional affiliations.
- Responsibilities: The editors, editorial board and publisher do not assume any responsibility for the article's contents and for the authors' views expressed in their contributions. The statements and opinions published represent the views of the authors or persons to whom they are credited. Publication of research information does not constitute a recommendation or endorsement of products involved.